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ANTIGENIC CONSTRUCTS OF MAJOR HISTOCOMPATIBILITY COMPLEX CLASS I ANTIGENS
WITH SPECIFIC CARRIER MOLECULES, THE PREPARATION AND USE THEREOF
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WO 80/01986
- (57) It has been found that target-cell-specific carriers, for example preferably monoclonal antibodies (mAb), but also polyclonal antibodies or molecules which bind to receptors on cells, can be coupled to the N- or C-terminal end of an allogenic MHC class I molecule without this altering the allogenic determinants in a disadvantageous manner. The MHC class I molecule is brought, with the aid of this target-cell-specific carrier, specifically to the target cells, which leads to activation of allospecific T cells and thus to destruction of the target cells by allospecific cytotoxic T cells. One explanation for the success of the coupling of a target-cell-specific carrier to the N- or C-terminal end of a MHC class I molecule while retaining the allogenic determinants is that the N-terminal end of the MHC class I molecule is located on the side of the α_1 and α_2 domains which points towards the cell, whereas the allogenic determinants are located on the side of the α_1 and α_2 domains which faces away from the cell (Fig. 1, Fig. 34).

CLAIM

1. Antigenic constructs adapted to induce the specific activation of allospecific T cells and in which major histocompatibility complex (MHC) class I antigens are linked at the C- or N- terminal end to specific carrier molecules.
7. Antigenic constructs as claimed in claim 6, in which the monoclonal antibodies are shortened in the constant part of the heavy chain.
8. Antigenic constructs as claimed in claim 1, 2, 3, 4, 5, 6 or 7, in which the MHC class I antigen is HLA B27w or HLA B27k.
12. A process for the preparation of antigenic constructs as claimed in claim 1, 2, 3, 4, 5, 6, 7, 8 or 9, which comprises the required parts of genes being fused in the form of their DNA, being provided with suitable regulation sequences and being expressed in suitable expression systems.
14. A pharmaceutical which contains antigenic constructs as claimed in any one of claims 1 to 11.

(CONVENTION. By one or more persons and/or a Company.) Form 4

625065

COMMONWEALTH OF AUSTRALIA

Patents Act 1952-1989

CONVENTION APPLICATION FOR A PATENT

(1) Here
insert (in
full) Name
or Names of
Applicant or
Applicants,
followed by
Address (es).

NI
We of BEHRINGWERKE AKTIENGESellschaft
D-3550 Marburg, Federal Republic of Germany

(2) Here
insert Title
of Invention.

hereby apply for the grant of a Patent for an invention entitled:
ANTIGENIC CONSTRUCTS OF MAJOR HISTOCOMPATIBILITY COMPLEX CLASS I
ANTIGENS WITH SPECIFIC CARRIER MOLECULES, THE PREPARATION AND USE
THEREOF

(3) Here insert
number(s)
of basic
application(s)

which is described in the accompanying complete specification. This application is a
Convention application and is based on the application numbered⁽³⁾

P38 25 615.0

(4) Here insert
Name of basic
Country or
Countries, and
date or
dates

for a patent or similar protection made in⁽⁴⁾ Federal Republic of Germany
on 28th July 1983

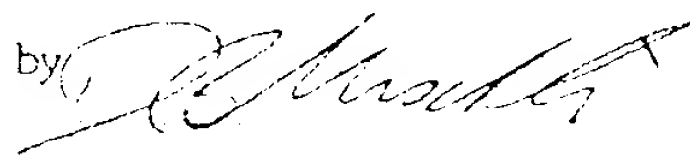
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DATED this 26th day of July 1989

BEHRINGWERKE AKTIENGESellschaft

(5) Signa-
ture (s) of
Applicant (s)
or
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authorized in
its Articles of
Association

27
G 11019 260789

by 
D. B. Mischlewski
Registered Patent Attorney

To:

COMMONWEALTH OF AUSTRALIAPatents Act 1952DECLARATION IN SUPPORT OF A CONVENTION APPLICATION UNDER PART XVI.
FOR A PATENT.

In support of the Convention application made under Part XVI. of the Patents Act 1952 by BEHRINGWERKE AKTIENGESELLSCHAFT of D-3550 Marburg, Federal Republic of Germany for a patent for an invention entitled: Antigenic constituents of raper and compatibility of class I antigens with specific carrier molecules, the preparation and use thereof

We, Philipp Stein, Eichenweg 76, D-3550 Marburg,
Heribert Bug, Amelweg 7, D-3550 Marburg,
Federal Republic of Germany

do solemnly and sincerely declare as follows:

1. We are authorized by BEHRINGWERKE AKTIENGESELLSCHAFT the applicant for the patent to make this declaration on its behalf.
2. The basic application(s) as defined by Section 141 of the Act was (were) made at München in the Federal Republic of Germany under No. P. A. 25 61510

on July 28, 1980

by BEHRINGWERKE AKTIENGESELLSCHAFT

3. a) Gerhard Seemann, Auf der Eber 1, D-3550 Marburg
b) Klaus Bosslet, Am Schlang 5, D-3550 Marburg
c) Hans Harald Sedlitz, Scherzhang 1, D-3550 Marburg
d) - d) Federal Republic of Germany

is/are the actual inventor(s) of the invention and the facts upon which BEHRINGWERKE AKTIENGESELLSCHAFT

is entitled to make the application are as follows:

The said BEHRINGWERKE AKTIENGESELLSCHAFT
is the assignee of the said

Gerhard Seemann, Klaus Bosslet, Hans Harald Sedlitz

4. The basic application referred to in paragraph 2 of this Declaration was (were) the first application(s) made in a Convention country in respect of the invention the subject of the application.

DECLARED at Marburg, Federal Republic of Germany

this 20th day of June 1980

To the Commissioner of Patents

BEHRINGWERKE AKTIENGESELLSCHAFT

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Prokurist

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ppa Bug
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Form 10

COMMONWEALTH OF AUSTRALIA

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COMPLETE SPECIFICATION

(ORIGINAL)

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Int. Class

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Complete Specification for the invention entitled:

ANTIGENIC CONSTRUCTS OF MAJOR HISTOCOMPATIBILITY COMPLEX CLASS I
ANTIGENS WITH SPECIFIC CARRIER MOLECULES, THE PREPARATION AND USE
THEREOF

The following statement is a full description of this invention, including the best method of performing it known to the applicant.

Antigenic constructs of major histocompatibility complex class I antigens with specific carrier molecules, the preparation and use thereof

5 The invention relates to antigenic constructs resulting from the linkage of major histocompatibility complex (MHC) class I antigens with specific carrier molecules.

10 Tissue-rejection reactions are the strongest-known immune responses mediated by T cells. In individuals of the same species they are caused by allogenic differences in class I and class II MHC antigens. In organ transplants, for example, any allogenic determinants of the MHC antigens present in the donor tissue are recognized as foreign by allospecific T cells of the recipient, a T cell immune response is induced, and the rejection reaction takes place unless an immunosuppressive therapy has been initiated or such a therapy proves insufficient.

15 It is furthermore known that MHC class I antigens are glycoproteins which are expressed on the surface of all nucleated cells. They are composed of a heavy chain, which is encoded by MHC class I genes, and of a light chain, the β_2 -microglobulin which is non-covalently associated with the heavy chain. The extracellular part of the heavy chain is folded in three domains, the first two of these domains (α_1 and α_2) exhibiting a pronounced polymorphism when the amino acid sequences of hitherto known class I MHC antigens from various individuals are compared. They assist with antigen presentation and carry the allogenic determinants. The third extracellular domain has a more conserved sequence. The association with β_2 -microglobulin is essential for correct folding of the heavy chain and for the transport of the molecule to the cell surface.

Isolation and characterization of mutated MHC class I antigens in mice showed that merely a few differences in amino acids on the α_1 and α_2 domains between donor and recipient suffice to induce a rejection reaction (Nathenson et al., Ann. Rev. Immunol., 1986, 4, 471-502). It has also been shown in humans that slight differences between donor and recipient lead to rejection of a transplant (Dausset, J., Rapaport, F.T., Legrand, L., Colombani, J., Marcelli-Barge, A.: Skin allograft survival in 238 human subjects: Role of specific relationships at the four gene sites of the first and the second HL-A loci., Histocompatibility Testing (1970) pages 381-397, Terasaki P.I. (Ed.)). The task which presented itself from that said above was to utilize the specific inducibility and strength of the cellular immune response in the tissue-rejection reaction to damage or destroy selected target cells.

It has been found that target-cell-specific carriers, for example preferably monoclonal antibodies (mAb), but also polyclonal antibodies or molecules which bind to receptors on cells, can be coupled to the N- or C-terminal end of an allogenic MHC class I molecule without this altering the allogenic determinants in a disadvantageous manner. The MHC class I molecule is brought, with the aid of this target-cell-specific carrier, specifically to the target cells, which leads to activation of allospecific T cells and thus to destruction of the target cells by allospecific cytotoxic T cells. One explanation for the success of the coupling of a target-cell-specific carrier to the N- or C-terminal end of a MHC class I molecule while retaining the allogenic determinants is that the N-terminal end of the MHC class I molecule is located on the side of the α_1 and α_2 domains which points towards the cell, whereas the allogenic determinants are located on the side of the α_1 and α_2 domains which faces away from the cell (Fig. 1, Fig. 34).

Because of the great polymorphism of the MHC class I

antigens in the human population, it is possible to induce a rejection reaction in almost 100 % of the population with the aid of only two different MHC class I molecules selected, for example HLA B27w and HLA B27k. HLA B27w and HLA B27k are two subtypes of the serologically defined HLA B27 specificity which are defined by cytotoxic T lymphocytes. In the caucasoid population about 7 % of individuals express HLA B27w and about 1 % express HLA B27k. The use of, for example, both HLA B27 subtypes for the allogenization in accordance with this invention makes it possible to treat almost 100 % of the caucasoid population. However, it is possible according to the invention to couple any desired MHC class I antigen to the relevant specific carriers if the above-mentioned antigens do not lead in the relevant recipient to activation of allospecific T cells and subsequent damage to or destruction of the target cells. Target cells may be regarded as cells which are undesired and/or pathogenic in the body, such as, for example, tumor cells. The antigenic constructs according to the invention are accordingly suitable for tumor therapy. However, it is also possible with the MHC class I antigenic constructs according to the invention to treat other diseases which are caused by cells or the products thereof and are favorably affected by elimination of these cells. The mode of action of the hybrid molecules described in example groups I and II derives from the fact that they are able, because of the specificity of the antibody portion, to bind to an antigen on the cell. The HLA B27 portion of the fused molecule results in masking of the surface of the target cell with an allogenic MHC class I molecule. These allogenic class I molecules can then be recognized by syngeneic, allospecific, cytotoxic T cells, which leads to destruction of the target cells by the allospecific cytotoxic T cells. Accordingly, the invention relates to

- a) MHC class I antigens which are linked N- or C-terminally to specific carriers, the linkage

preferably being brought about covalently but also possibly being non-covalent, for example by a biotin-avidin bridge, and the specific carriers binding selectively to target cells and denoting preferably monoclonal, but also polyclonal, antibodies, but being very generally receptor-binding molecules which bind to the particular cell receptors,

b) a process for the preparation of the MHC class I antigenic constructs, and

c) the use of the MHC class I antigenic constructs mentioned in a) and b) for damaging or eliminating target cells.

The invention is furthermore described in the examples which follow and in the patent claims, but it is not to be regarded as restricted thereto.

Examples 1 - 17 detailed hereinafter describe a construct according to the invention composed of the nitrophenol (NP)-specific mouse mAb B/1-8 V_H gene (1), of a human IgG C F(ab')₂ gene (2) and of an HLA B27w gene (3). (1) and (2) are to be regarded in this context as examples of the specific carrier portion - in this case an mAb against NP - whereas (3) represents an HLA class I antigen.

The abovementioned construct is, after appropriate transformation, expressed and secreted by those myeloma cells which contain a human β_2 -microglobulin and a light chain of an immunoglobulin and whose V gene forms with V_H B/1-8 a NP binding-site, such as, for example, the mice myeloma cell J 558 L (Oi, V.T., Morrison, S.L., Herzenberg, L.A., Berg, P.: Immunoglobulin gene expression in transformed lymphoid cells. Proc. Natl. Acad. Sci. USA 80, 825, 1983). It is possible, by exchanging the V_H gene of the heavy chain and using an appropriate light chain, to provide the mAb/HLA B27w fusion product

with any desired specificity for which a specific or selective mAb exists.

Examples

I Examples 1 to 13 show the construction of an HLA
5 B27/mAb fusion gene with the HLA B27 portion at the
 3' end of the monoclonal antibody

A) Preparation of the mAb C gene portion (IgG₃ C gene)

Example 1

10 A human IgG₃ C gene was isolated from a human gene bank
 in EMBL3 phages (Frischauf, A.-M., Lehrach, H., Proustka,
 A., Murray, N.: Lambda replacement vectors carrying poly-
 linker sequences. J. Mol. Biol. 170, 827-842 (1983) and
 Seemann, G.H.A., Rein, R.S., Brown, C.S., Ploegh, H.L.:
15 Gene conversion-like mechanisms may generate polymorphism
 in human class I genes. The EMBO Journal 5, 547-552
 (1986)) and subcloned as a HindIII/Sph I fragment 3.1 kb
 in size into the plasmid vector pUC 19 (clone 54.1.24)
 (Fig. 2).

20 All the techniques used in these and in the following
 examples were taken, unless otherwise indicated, from
 Lehrach, H. and Frischauf, A.-M. Laboratory Manual EMBL
 (1982), Heidelberg; Maniatis, T., Fritsch, E.F.,
 Sambrook, J.: Molecular Cloning: A laboratory manual
 (1982), Cold Spring Harbor Laboratory.

25 Example 2

 The 54.1.24 clone was subjected to complete HindIII and
 partial PstI restriction digestion. This results, inter
 alia, in restriction fragments which contain the C_{H1} exon
 and one, two or three hinge exons. These fragments were
30 cut out of an agarose gel and cloned into a pUC 19 vector
 cut with HindIII and PstI (Fig. 3).

The plasmid clone with the C_{H1} and three hinge exons ($F(ab')_2$ 3H) was then cleaved with BamHI and Asp 718, the cleavage sites were filled in and religated with T_4 ligase (Fig. 4). This deletes the pUC 19 polylinker between the Xba I and the SstI cleavage site.

I B) Preparation of the HLA B27 gene

Example 3

An HLA B27w gene was isolated from a genomic gene bank cloned in EMBL3 bacteriophages (Frischauf, A.-M., loc. cit., and Seemann, G.H.A., loc. cit.) and characterized by restriction mapping and nucleotide sequence analysis (Maxam, A., Gilbert, W.: Sequencing end-labeled DNA with base specific chemical cleavage. Meth. Enzymol. 65, 499-560 (1980) and Sanger, F., Nicklen, S., Coulson, A.R.: DNA sequencing with chain terminating inhibitors. Proc. Natl. Acad. Sci. USA 74, 5463-5471 (1977)) (Fig. 5).

The HLA B27w gene was then digested with the restriction enzymes SstI and BglII and subcloned into the SstI and BamHI cleavage sites of pUC 19. Plasmid clones with the subfragments A, B and C (Fig. 5) were isolated.

Example 4

The plasmid with subfragment A was cleaved completely with SstI and partially with SmaI and, after fractionation on an agarose gel, the fragment A' (Fig. 6) was cloned in a pUC 19 plasmid cleaved with HincII and SstI.

Example 5

The plasmid with the subfragment B was digested with XbaI, and the resulting XbaI insert (B') was cloned in a XbaI-cleaved pUC 19 plasmid (Fig. 7).

Example 6

5 The plasmid with the subfragment C was cleaved completely with HindIII and partially with SstI, and the fragment which, in the HLA B27w gene, is attached to fragment A (C') was, after fractionation on an agarose gel, isolated and cloned into the Bluescript KS+ phasmid vector (Stratagene, LaJolla, CA, USA) cleaved with HindIII and SstI (Fig. 8).

Example 7

10 Single-stranded phages were prepared from the KS+ phasmid vector C' by infection with VCS-M13 helper phages (Stratagene, Cat # 200251) and were purified (Stratagene: Bluescript Exo/Mung DNA sequencing system: Instruction Manual). A synthetic oligonucleotide (I =
15 5'CCTTACCTCATCTCAGG3') was hybridized onto these single strands, and the remainder of the second strand was synthesized using Klenow polymerase. The double-stranded phasmids generated in this way were transformed into XL Blue bacteria and then single-stranded phages were again
20 generated from the resulting plasmid clones by infection with helper phages, and the nucleotide sequence was determined with the aid of an oligonucleotide primer II (5'TGAGGGCTCCTGCTT3') (Sanger, F. et al., loc. cit.). A clone in which the codon TGG (amino acid 274) at the 3'
25 end of the alpha3 exon had been mutated to a stop codon (TGA) was identified (C'') (Fig. 9).

Example 8

30 The plasmid with the fragment A' was cleaved with SstI and ligated with the C'' fragment which had been generated by a complete HindIII and partial SstI cleavage of the phasmid clone C'' and had been isolated after fractionation on an agarose gel. After ligation at 14°C for 30 minutes, the unligated ends were filled in with T₄ polymerase and subsequently ligated once again. Restriction mapping was

used to identify the plasmid D (Fig. 10) in which the fragment A' is connected to the fragment C" via the SstI cleavage site in the alpha2 exon.

Example 9

5 The plasmid with the fragment D was cleaved with XbaI and ligated with the fragment B' which had been cut out of the plasmid B' with XbaI and had been purified after fractionation on an agarose gel (Fig. 11). Nucleotide sequence analyses (17) were used to identify a plasmid
10 (E) in which the B' fragment is ligated in the correct 5'-3' orientation to the fragment D.

C) Fusion of the modified HLA B27w gene with the IgG3 C F(ab')₂ 3H gene fragment

Example 10

15 The fragment E was cut out of the plasmid E by cleavage with EcoRI and HindIII, the ends were filled in with T₄ polymerase and purified after fractionation on an agarose gel. This purified fragment E was then ligated with the plasmid which contains the IgG3 F(ab')₂ 3H fragment after
20 the latter had been cleaved and the XbaI ends had been filled in with T₄ polymerase (Fig. 12). Restriction mapping was used to identify the clone which contained the plasmid F, in which the modified HLA B27w gene is fused in the correct 5'-3' orientation to the F(ab')₂ 3H gene.
25

Example 11

The fragment F was cut out with HindIII and EcoRI in order to place a polylinker in front of the 5' end of the fragment F. The HindIII and XbaI ends were filled in with
30 T₄ polymerase and cloned into a pUC 19 which had been cleaved with SstI and whose SstI ends had been filled in with T₄ polymerase. Restriction analyses were used to

identify the clone with the plasmid G which has the pUC 19 polylinker 5' from the fragment F (Fig. 13).

Example 12

The plasmid G was cleaved with HindIII and EcoRI, and the insert with the IgG F(ab')₂ HLA B27w fusion gene was isolated and cloned into a Bluescript KS+ phasmid vector (Stratagene: Bluescript Exo/Mung DNA sequencing system: Instruction Manual) cleaved with HindIII and EcoRI (Fig. 14).

Example 13

The plasmid H resulting from this cloning was then cleaved with BamHI, and the insert was cloned into the eukaryotic expression vector pEV_E (Simon, T., Rajewsky, K., Nucl. Acids Res. 16, 354, (1988), which contains the IgG H promoter/enhancer sequences and the V_E gene originating from the NP-specific mouse mAb B/1-8 which had been cleaved with BamHI (Fig. 15) (Neuberger, M.N.: EMBO Journal 2, 1375-1378 (1983)). Restriction analysis was used to identify the plasmid I in which the IgG 3 F(ab')₂ HLA B27w fusion gene is cloned in the correct 5'-3' orientation behind the V_E gene.

The mAb/HLA B27w fusion gene now possesses intact 5' and 3' ends having all the signals required for expression in eukaryotic cells. The construct is, as stated in the introduction, expressed and secreted in every myeloma cell which contains a human β_2 -microglobulin and a light chain of an immunoglobulin and whose V gene forms with V_E B/1-8 a NP binding site, such as, for example, the mouse myeloma cell J 558L (Oi, V.T., Morrison, S.L., Herzenberg, L.A., Berg, P., Proc. Natl. Acad. Sci. USA 80, 825 (1983)).

II Examples 14 to 17 show the construction of an HLA B27/mAb fusion gene with the HLA B27 portion at the

5' end of the monoclonal antibody

A) Preparation of the HLA B27 gene

Example 14

5 An HLA B27w gene was isolated from a genomic gene bank cloned in EMBL3 bacteriophages (Frischauf et al., loc. cit. and Seemann, G.H.A., loc. cit.) and characterized by restriction mapping and nucleotide sequence analysis (Maxam et al., loc. cit. and Sanger et al., loc. cit.) (Fig. 5).

10 The HLA B27w gene was then digested with the restriction enzymes SstI and BglII and subcloned into the SstI and BamHI cleavage sites of pUC 19. Plasmid clones with the subfragments A, B and C (Fig. 5) were isolated.

15 The plasmid with the HLA B27 subclone C was partially cleaved with PstI. The protruding 3' ends of the PstI cleavage sites were removed with T₄ polymerase, with addition of dGTP, and religated with T₄ ligase. Restriction analysis was used to identify the plasmid clone C1 which contains no PstI cleavage site in the intron
20 between the alpha2 and alpha3 exon (Fig. 16).

The plasmid clone C1 was cleaved partially with SstI and completely with HindIII, and the C1' fragment was isolated and cloned into a double-stranded M13 mp18 vector cleaved with SstI and HindIII. The M13 clone C1' with the
25 C1' fragment was identified by determining the nucleic acid sequence of the insert (Fig. 17).

Using the protocol of the Bio-Rad Muta-Gene M13 mutagenesis kit there were isolated from the C1' M13 mp18 phages in the bacterial strain CJ236 single-stranded
30 phages which contained uracils. An oligonucleotide (oligonucleotide III) with the sequence 5' GCGCGCTGGAGCGTCTC^{3'} was hybridized onto these single-stranded phages and the

second strand was synthesized with addition of T. polymerase, dNTP and T. ligase.

5 After infection of the bacterial strain MV 1190 the mutated clone C2 was identified by restriction analysis of the M13 mp18 double-stranded DNAs and confirmed by nucleic acid sequence analysis (Fig. 18). The mutagenesis resulted in destruction of the PstI restriction cleavage site in the alpha2 exon without altering the reading frame or the encoded amino acid sequence.

10 Single-stranded phages were in turn produced from the M13 clone C2 in the bacterial strain CJ236 and were hybridized with the oligonucleotide IV (oligonucleotide IV = 5'GGGGACGGTGGGAATTCGAAGACGGCTC3'). The second strand was then synthesized with T. polymerase, T. ligase and dNTP.
15 After transformation into MV 1190 bacteria, the M13 mp18 clone C2' was identified by restriction analysis, and the mutation was verified as correct by nucleotide sequence analysis (Fig. 19). This mutagenesis resulted in an EcoRI and an AsuII cleavage site being introduced into the TM exon of the HLA B27 gene, and in amino acid 279 being converted from glutamine into asparagine (Fig. 20).

Example 15

25 The plasmid clone with the subfragment B was digested with XbaI, and the resulting XbaI insert (B') was cloned in an XbaI-cleaved pUC 19 plasmid (Fig. 7).

30 The plasmid clone with the fragment A was cleaved completely with HindII and partially with SmaI, and religated. Restriction analysis was used to identify the clone A' in which part of the pUC 19 polylinker is deleted (Fig. 21).

The plasmid clone A' was cleaved partially with SstI and ligated with the C2 fragment generated by an SstI cleavage of the plasmid clone C2 and isolated after

fractionation on an agarose gel. Restriction mapping was used to identify the plasmid D₁ (Fig. 22) in which the fragment A is connected to the fragment C2 via the SstI cleavage site in the alpha2 exon. The 5' end of the HLA B27w gene is thus complete.

Construction of the linker:

Two oligonucleotides were synthesized:

oligonucleotide Va:

5' TCGAATTCCG GCGAGGCAGC TCCCGCAGCT GCACCCGCAG CAGCCGCAGC
AGGCGGGCAG GTCCAACCTGC AGGA 3'

oligonucleotide Vb:

5' TCCTGCAGTT GGACCTGCCC GCCTGCTGCG GCTGCTGCGG GTGCAGCTCC
GGGAGCTGCC TCGCCGGAAT TCGA 3'

The two oligonucleotides were hybridized together. This resulted in double-stranded DNA fragments with an EcoRI restriction cleavage site at one end and a PstI restriction cleavage site at the other end. These fragments were cleaved with EcoRI and PstI and cloned into an EcoRI- and PstI-cleaved pUC 19 plasmid vector (Fig. 23). The plasmid clone L was identified by restriction analysis and verified by nucleotide sequence analysis.

The immunoglobulin V gene was synthesized by P.T. Jones et al. (Jones, P.T., Dear, P.H., Foote, J., Neuberger, M.S. Winter, G., Nature 321: 522, (1986)) using oligonucleotides. It contains a PstI restriction cleavage site in the 5' region of the clone and is cloned as HindIII/PamHI fragment in an M13 mp8 vector whose PstI cleavage site had been destroyed by cleavage, removal of the protruding ends and religation (Fig. 24).

II B) Preparation of the mAb C gene portion:

Example 16

A human IgG3 C gene was isolated from a human gene bank in EMBL3 phages (Frischauf et al., loc. cit. and Seeman et al., loc. cit.) and subcloned into the plasmid vector pUC 19 as a HindIII/SphI fragment 3.1 kb in size (clone 54.1.24) (Fig. 2).

The plasmid clone 54.1.24 was cleaved with HindII and Asp718, the protruding ends of the Asp718 cleavage site were removed with T₄ polymerase and religated with T₄ ligase. Restriction analysis and nucleic acid sequence determination were used to identify the clone 54.1.24 Delta Pol which, apart from SphI, PstI, SstI and EcoRI, no longer contains any restriction cleavage sites 3' of the human IgG3 C gene (Fig. 25).

The plasmid clone 54.1.24 Delta Pol was digested with BglII and SphI. The protruding ends were removed with T₄ polymerase and religated with T₄ ligase. Restriction analysis and nucleic acid sequence determination were used to identify the clone I which now contains only the CH₁ exon of the human IgG3 C gene (Fig. 26).

The plasmid clone I was cleaved with PstI, and the protruding ends were removed with T₄ polymerase. The B' insert which had been cut with XbaI and filled in with T₄ polymerase to give blunt ends was ligated into the resulting blunt ends. Restriction analysis and nucleic acid sequence determination were used to identify the clone K (Fig. 27) which contains a human IgG₃C_{H1} exon and a 3' end of a HLA class I gene.

The plasmid clone K was cleaved with HindIII and EcoRI, the protruding ends were removed, and the insert was ligated in an SstI-cleaved pUC 19 plasmid whose ends had likewise been made blunt. The clone L which harbors the polylinker of the pUC 19 vector 5' from the C_{H1} exon was

identified (Fig. 28).

The plasmid clone L was cleaved with EcoRI and HindIII, and the insert was purified and ligated into a HindIII- and EcoRI-cleaved KS⁺ vector (Stratagene; Bluescript Exo/Mung DNA Sequencing System) whose PstI cleavage site had previously been destroyed by cleavage with PstI, T, polymerase treatment and religation. The clone M, from which it is possible to cut out the human C₈1 exon with the HLA class I 3' end by a Bam HI cleavage, was identified (Fig. 29).

Example 17

Double-stranded DNA was prepared from the M13 mp8 clone V and cleaved with BamHI. The KS⁺ clone M was cleaved with BamHI, and the insert M was purified. The M fragment was ligated into the BamHI-cleaved clone V, and nucleic acid sequence determination was used to identify the M13 clone N which contains an intact IgG3 gene (Fig. 30).

Double-stranded DNA was prepared from the M13 clone N and was cleaved with EcoRI, and the insert was purified. The plasmid clone D₁ was cleaved with EcoRI and ligated with the fragment N. The phage clone O in which the fragment N is cloned in the correct orientation into the clone D₁ was isolated (Fig. 31).

The plasmid clone O was subjected to a complete PstI cleavage and partial EcoRI cleavage and ligated with the linker fragment cut out of the plasmid vector L with EcoRI and PstI. The plasmid clone P contains the complete HLA B27w mAb fusion gene (Fig. 32, Fig. 33). This fusion gene can be expressed and secreted in human cells alone or in mouse cells together with the human beta₂ microglobulin gene if the expressing cells also contain an immunoglobulin light chain.

Key to Fig. 1

5 alpha 1, alpha 2 and alpha 3 denote the domains of the class I MHC antigen chain. The arrows point to the alpha helices which carry the allodeterminants. CM is meant to represent the cell membrane, and C the cell.

Key to Fig. 2 et seq.

10 EcoRI etc. represents the cleavage with the particular restriction endonuclease or represents the corresponding cleavage site. BamHI/Asp 718 denotes a restriction cleavage site destroyed by religation after filling-in.

TM denotes the transmembrane region.

3'NT denotes 3' non-translated

IgH p/E denotes the immunoglobulin heavy chain promoter/enhancer

15 * denotes: incomplete digestion

DS-DNA denotes: double-stranded DNA

SS-DNA denotes: single-stranded DNA

Key to Fig. 34:

20 s.CTL denotes: syngeneic cytotoxic T lymphocyte

TcR denotes: T-cell receptor

a.MHC class I denotes: allogenic MHC class I antigen

t.a.a. denotes: tumor-associated antigen

s.t.c. denotes: syngeneic tumor cell



THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:

1. Antigenic constructs adapted to induce the specific activation of allospecific T cells and in which major histocompatibility complex (MHC) class I antigens are linked at the C- or N- terminal end to specific carrier molecules.
2. Antigenic constructs adapted to induce the specific activation of allospecific T cells and in which major histocompatibility complex (MHC) class I antigens are linked at the amino-terminal end to specific carrier molecules.
3. Antigenic constructs adapted to induce the specific activation of allospecific T cells and in which major histocompatibility complex (MHC) class I antigens are linked at the C-terminal end to specific carrier molecules.
4. Antigenic constructs as claimed in claim 1, 2 or 3, in which one MHC class I antigen is linked to one specific carrier molecule respectively.
5. Antigenic constructs as claimed in claim 1, 2, 3 or 4, in which the specific carrier molecules are CD4 domains.
6. Antigenic constructs as claimed in claim 1, 2, 3 or 4, in which the specific carrier molecules are monoclonal antibodies.
7. Antigenic constructs as claimed in claim 6, in which the monoclonal antibodies are shortened in the constant part of the heavy chain.
8. Antigenic constructs as claimed in claim 1, 2, 3, 4, 5, 6 or 7, in which the MHC class I antigen is HLA B27w or HLA B27k.
9. Antigenic constructs as claimed in claim 1, 2, 3, 4, 5, 6, 7 or 8, in which the MHC class I antigen is covalently bonded to the carrier molecule.
10. Antigenic constructs as claimed in claim 1, 2, 3, 4, 5, 6, 7 or 8, in which the MHC class I antigen is bonded



via avidin/biotin to the carrier molecule.

11. Antigenic constructs as claimed in claim 1, 2, 3, 4, 5, 6, 7 or 8, which are prepared by genetic manipulation by fusion of the DNAs coding for them.

12. A process for the preparation of antigenic constructs as claimed in claim 1, 2, 3, 4, 5, 6, 7, 8 or 9, which comprises the required parts of genes being fused in the form of their DNA, being provided with suitable regulation sequences and being expressed in suitable expression systems.

13. The use of the antigenic constructs as claimed in any one of claims 1 to 11 for the allogenization of target cells.

14. A pharmaceutical which contains antigenic constructs as claimed in any one of claims 1 to 11.

DATED this 26th day of July 1989
BEHRINGWERKE AKTIENGESELLSCHAFT

WATERMARK PATENT & TRADEMARK ATTORNEYS
50 QUEEN STREET
MELBOURNE, VIC. 3000.

FIG. 1

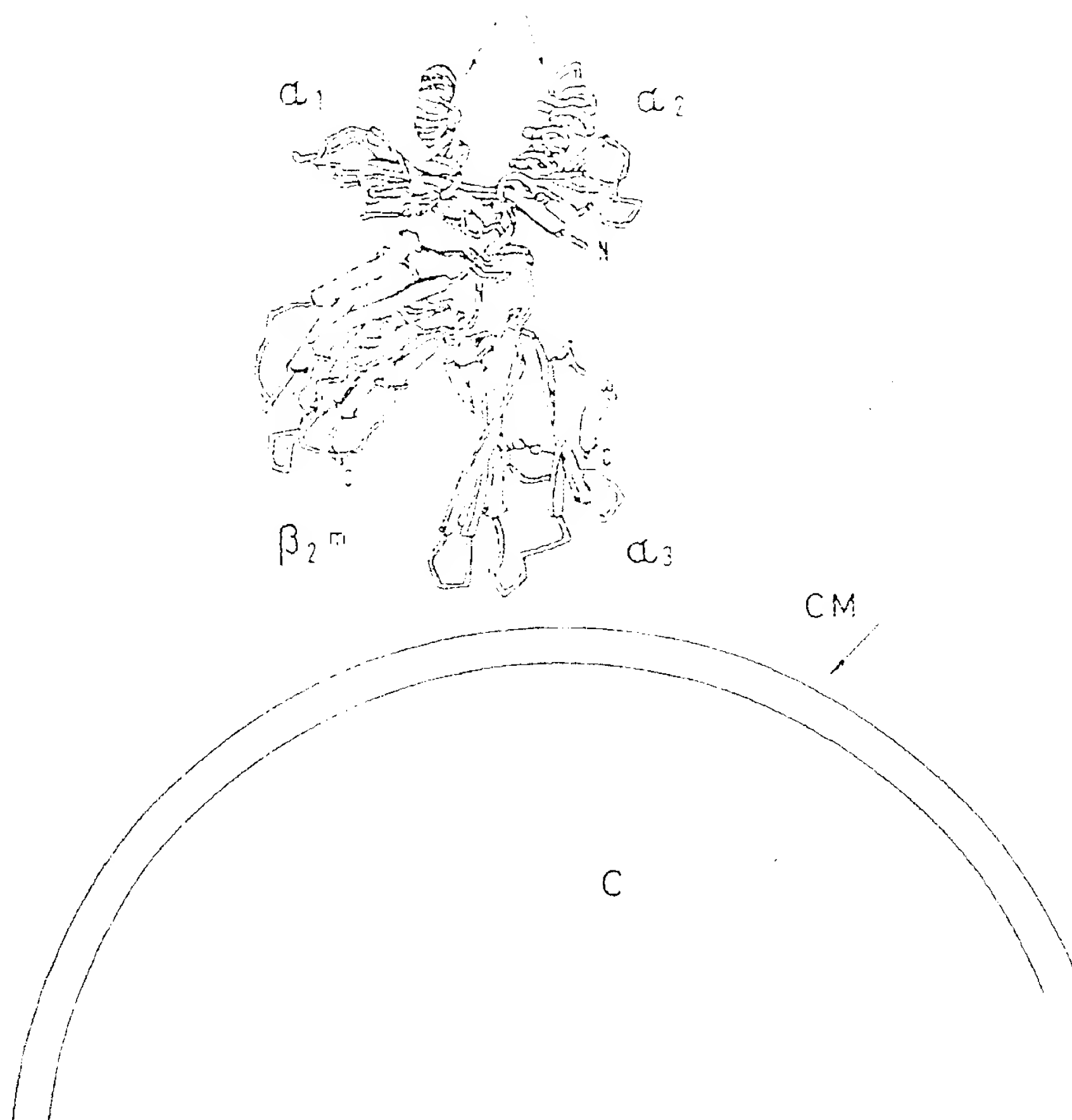
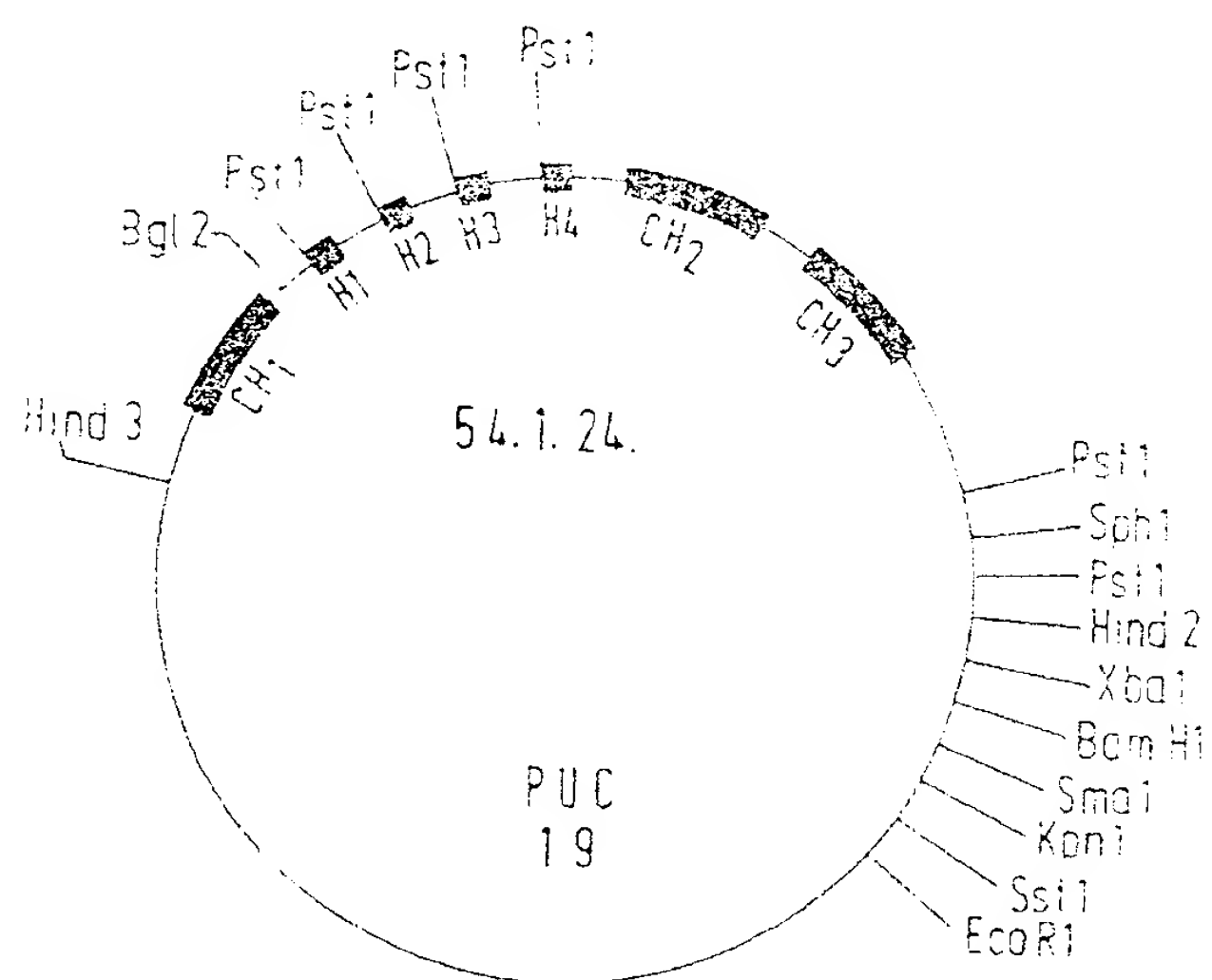


FIG. 2



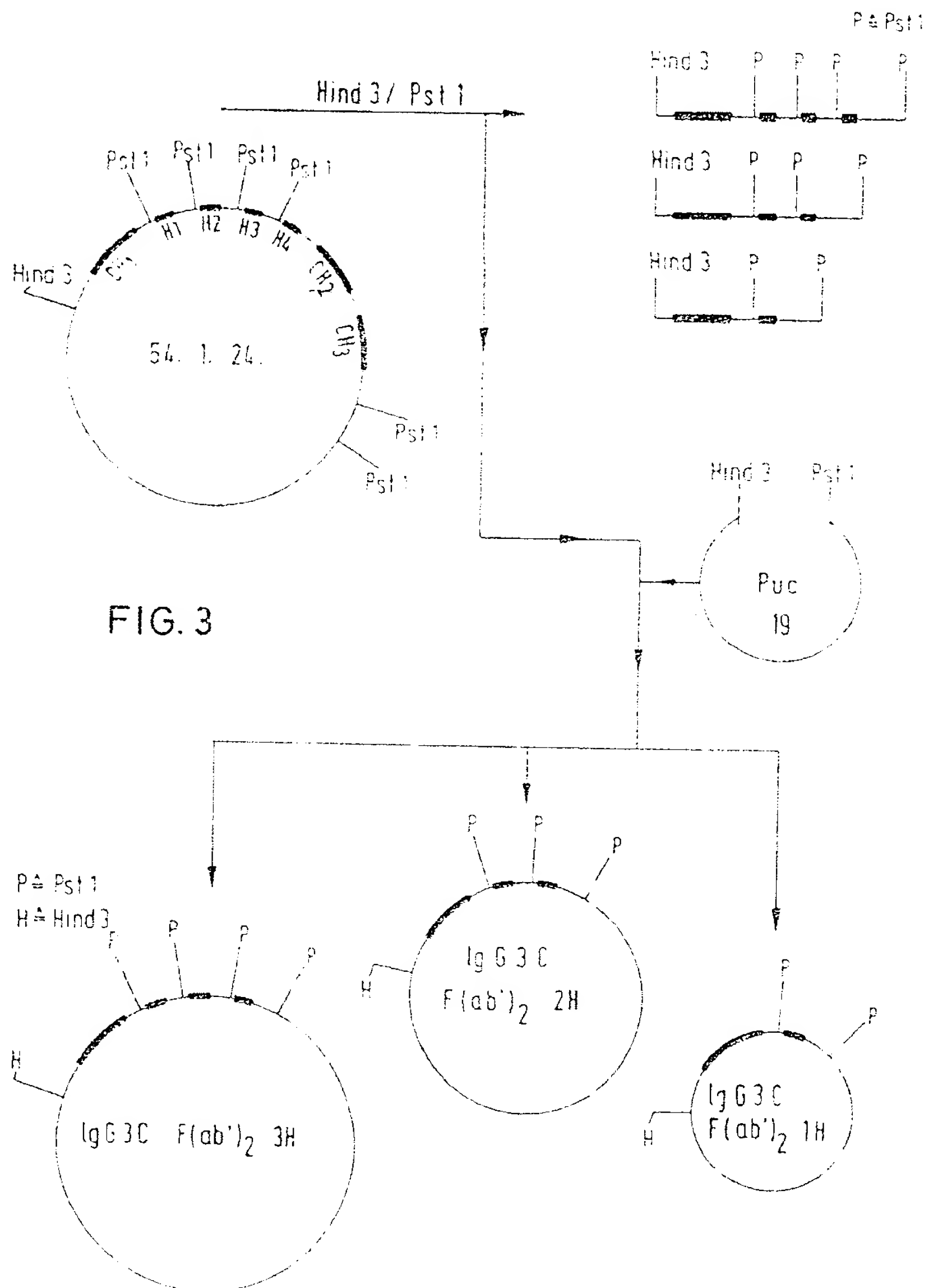
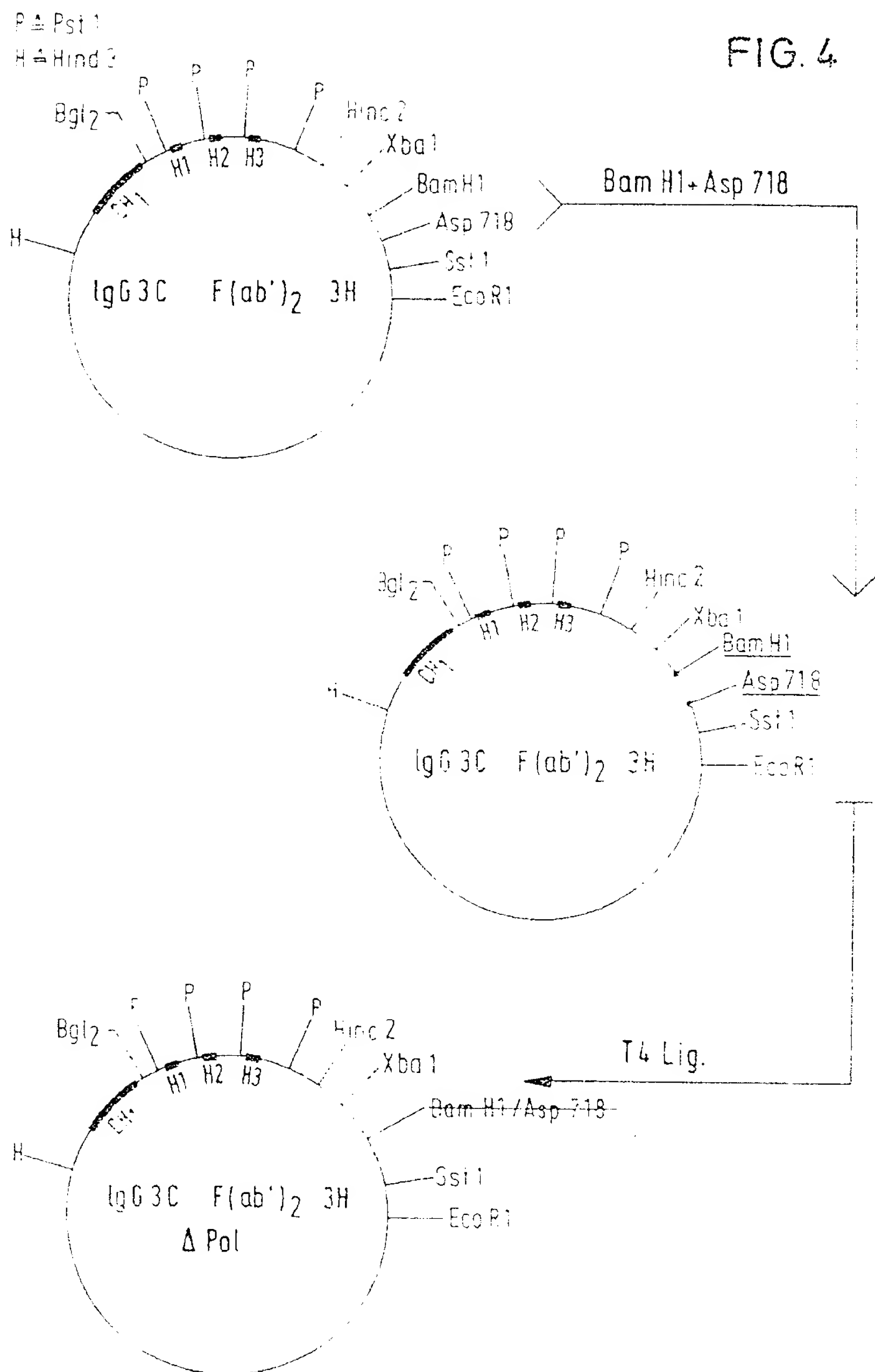
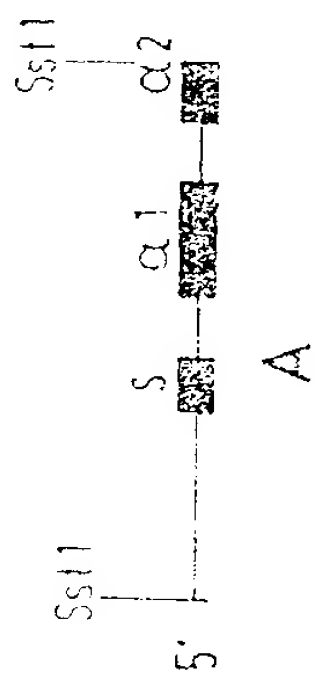
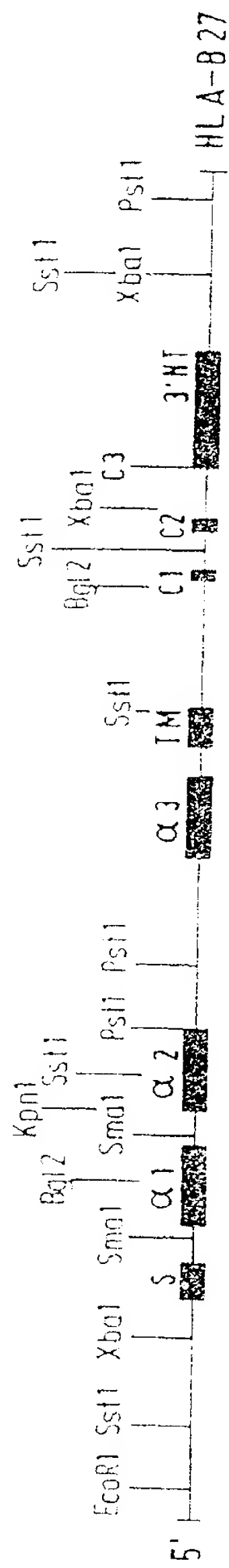
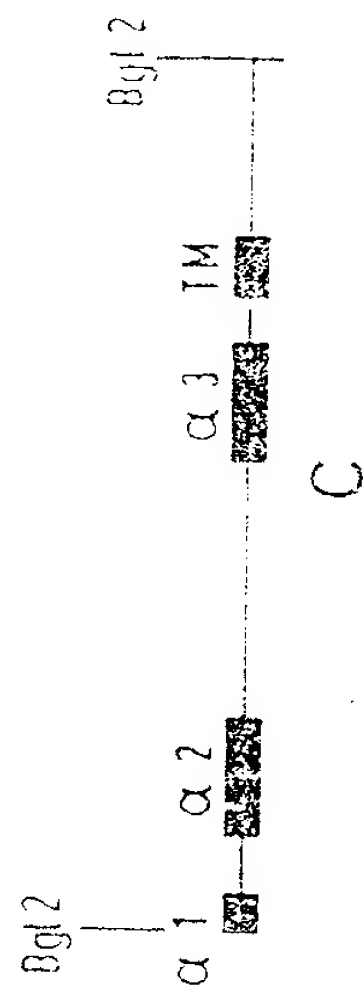


FIG. 3

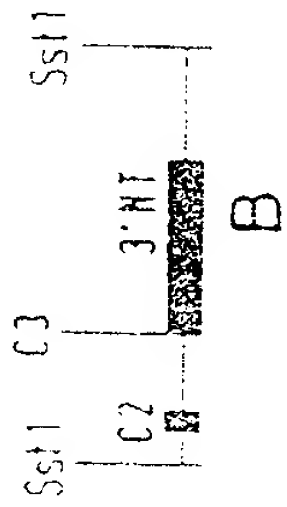




A



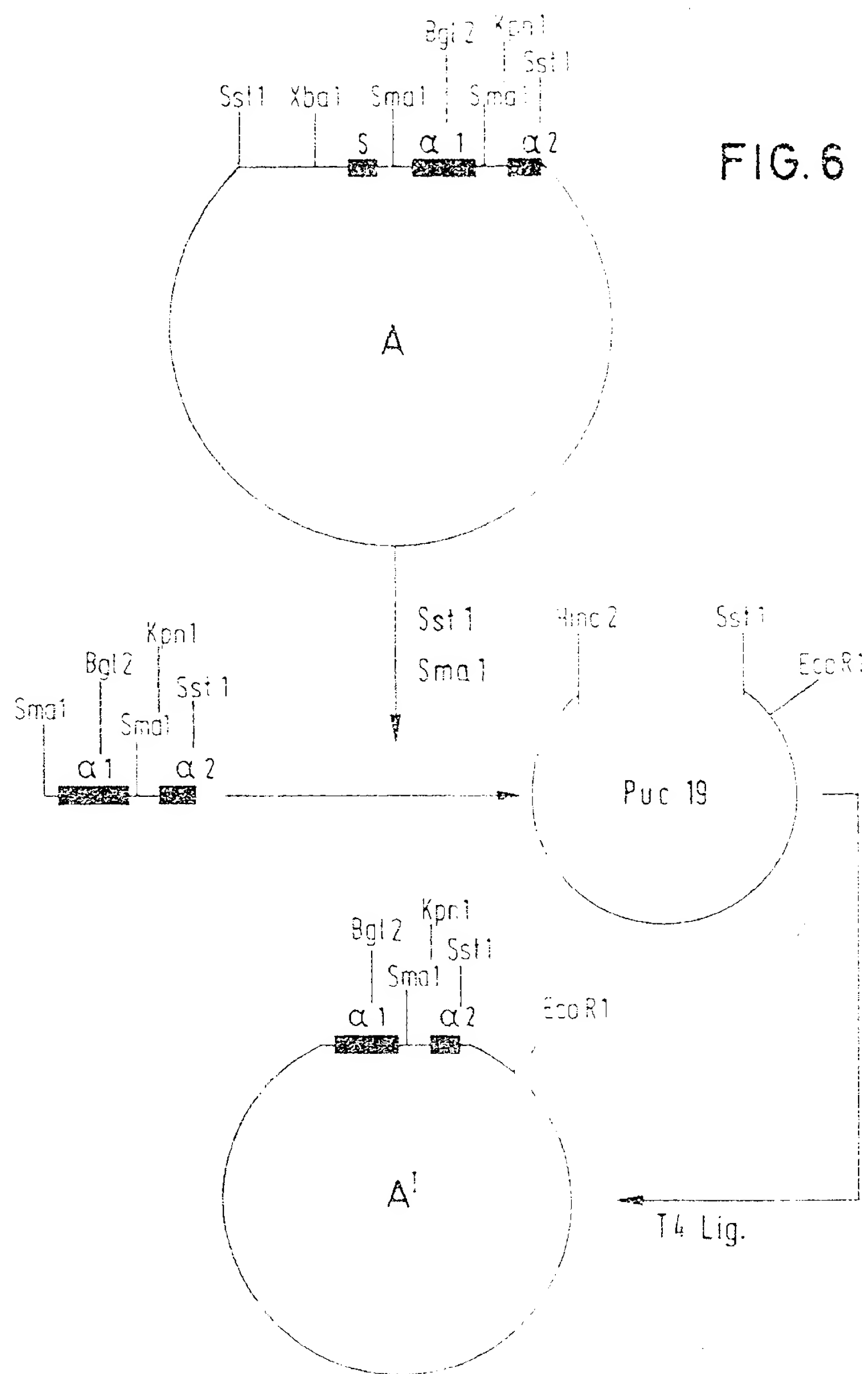
C



B

100 bp

FIG. 5



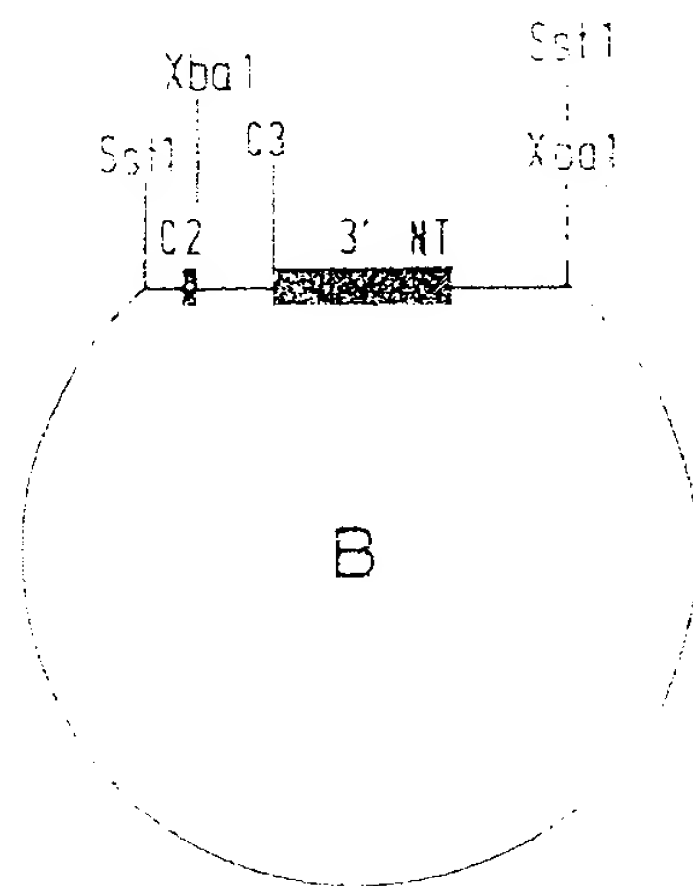
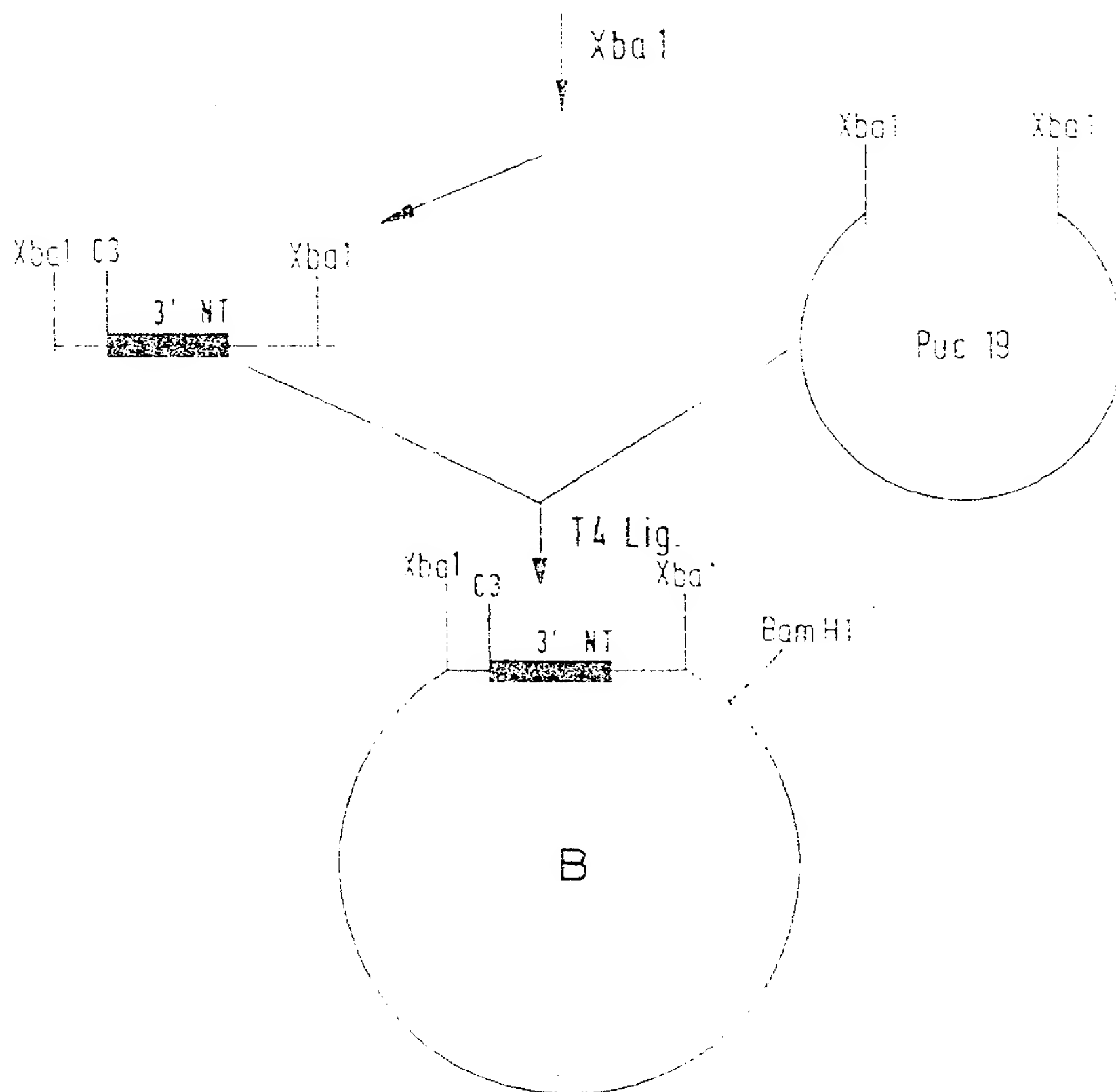
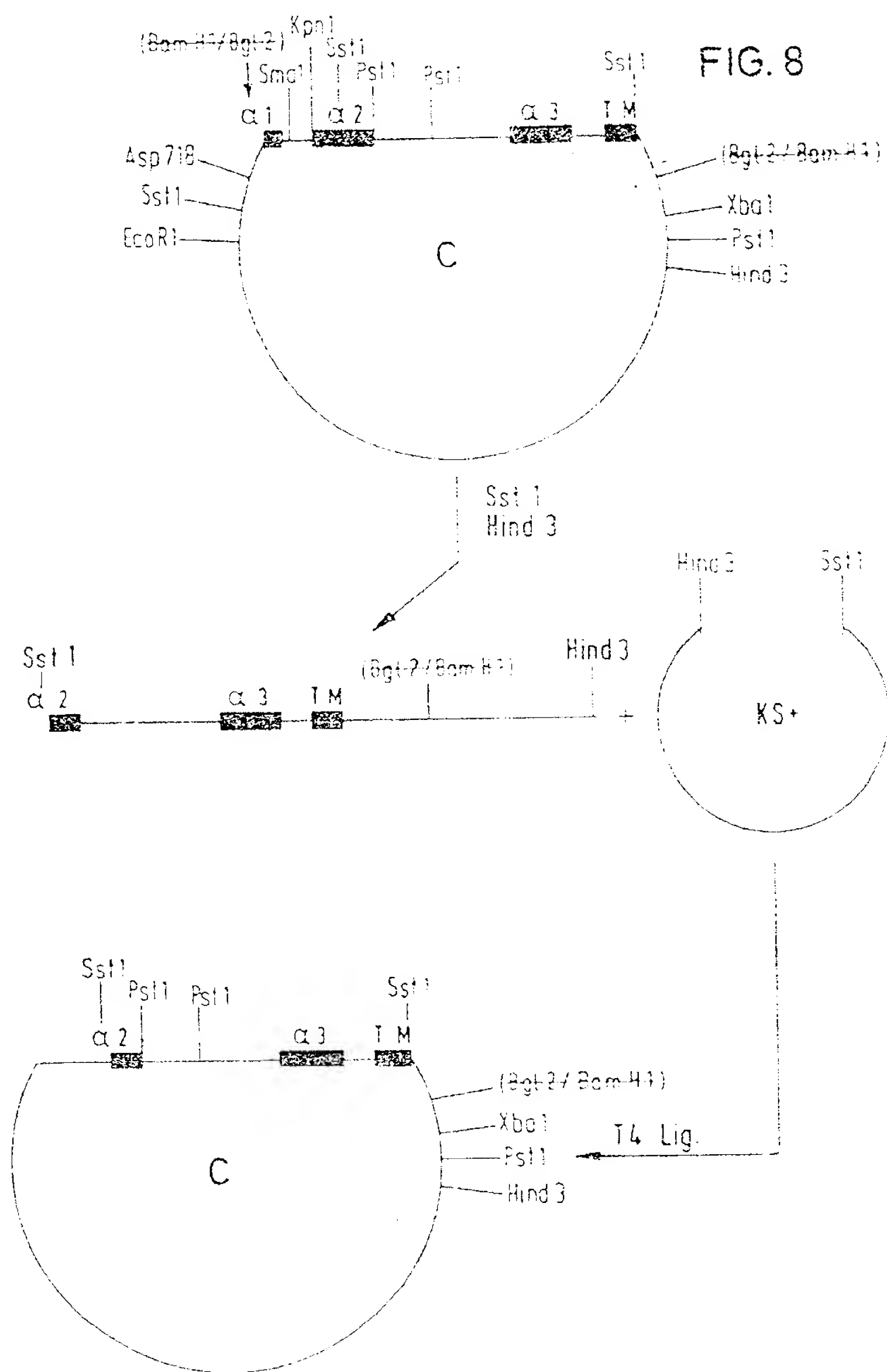


FIG. 7





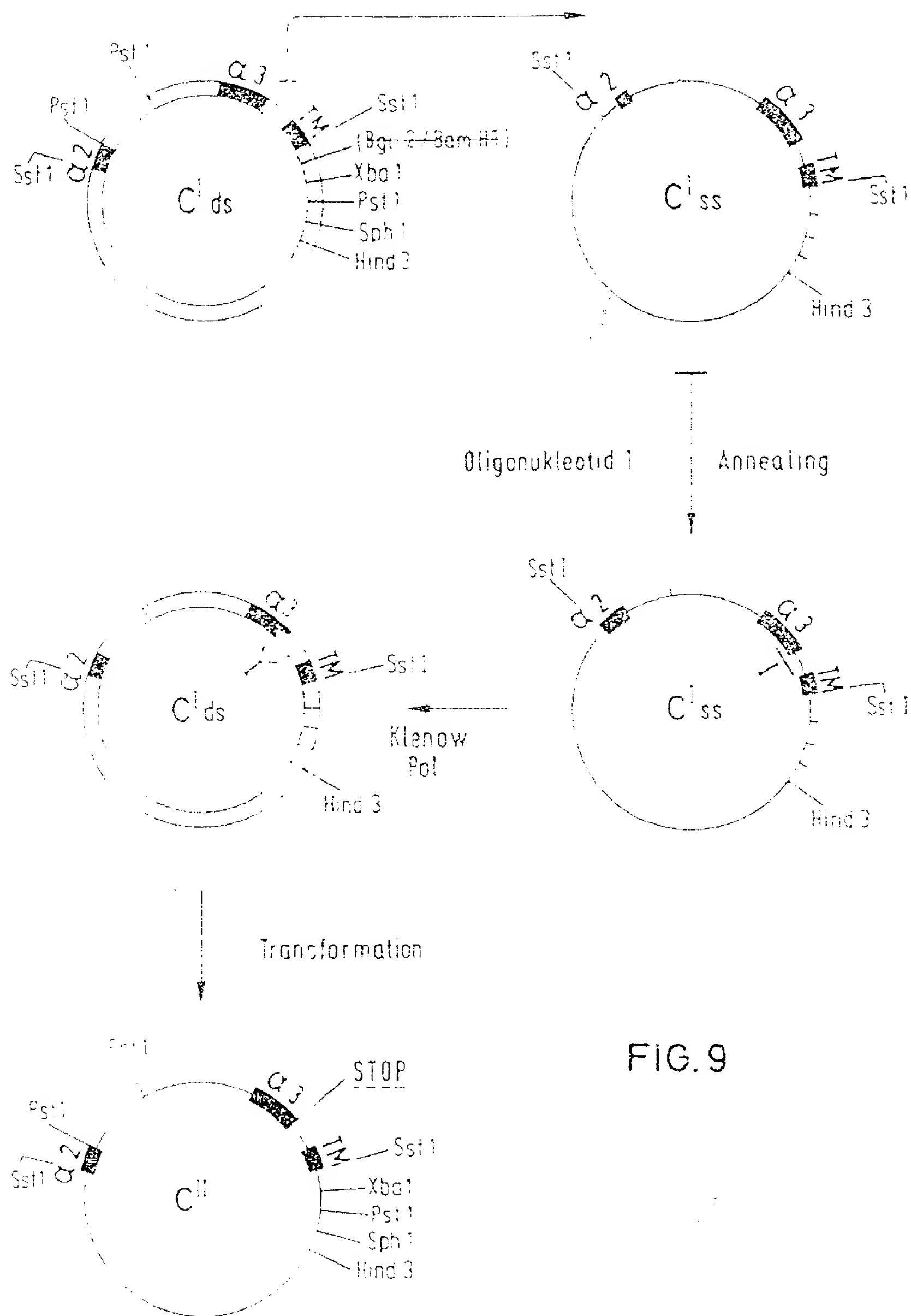


FIG. 9

FIG. 10

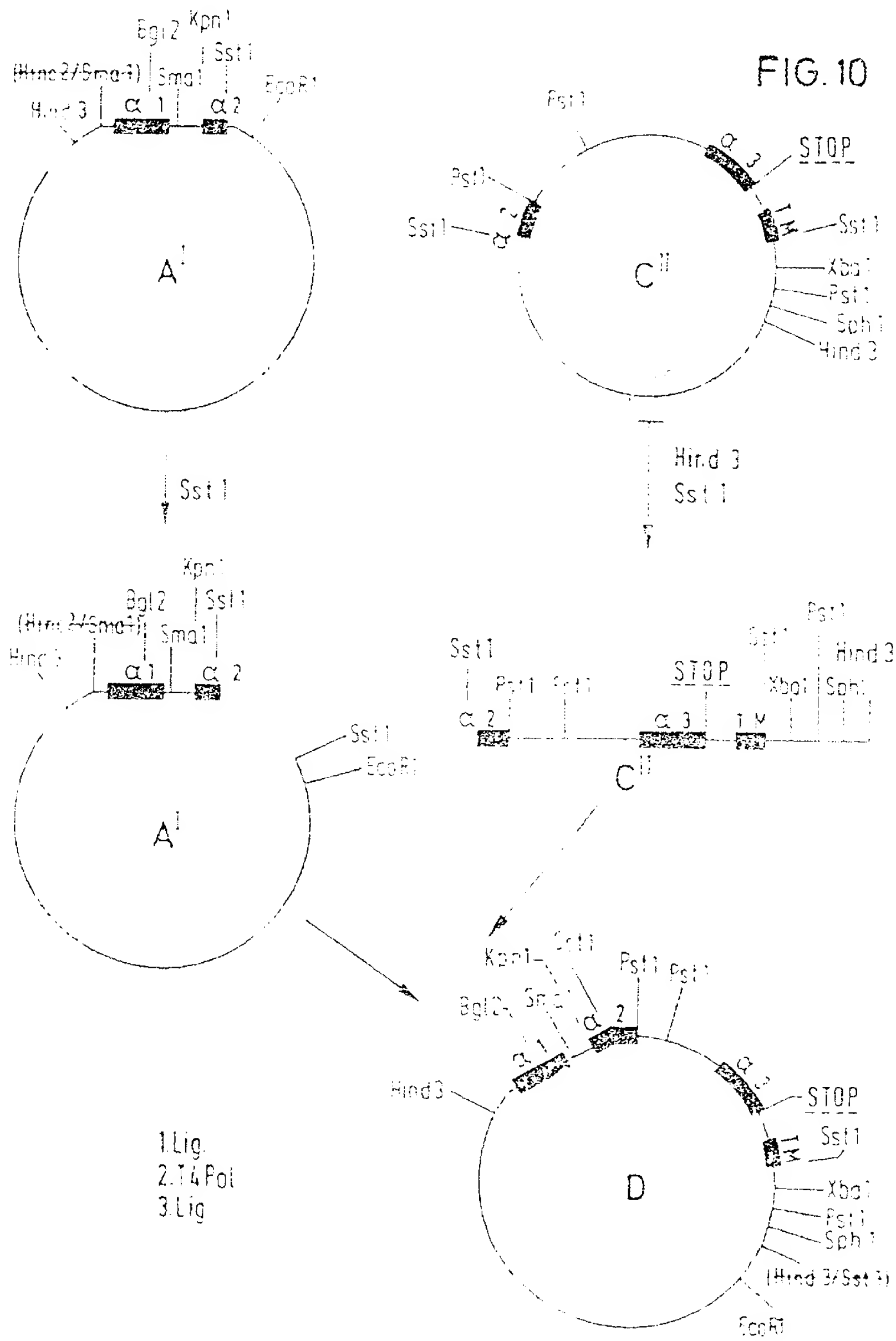
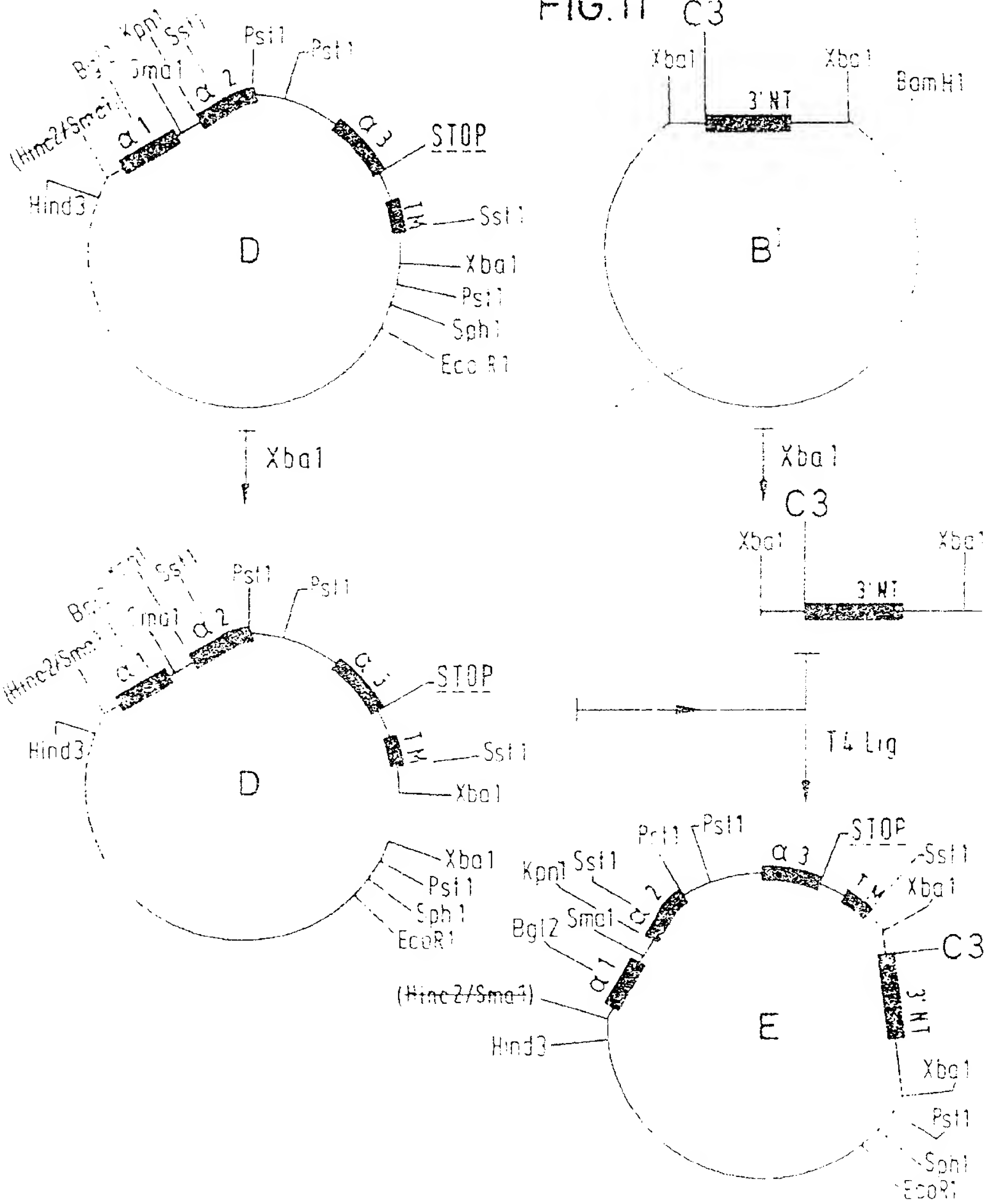


FIG.11 C3



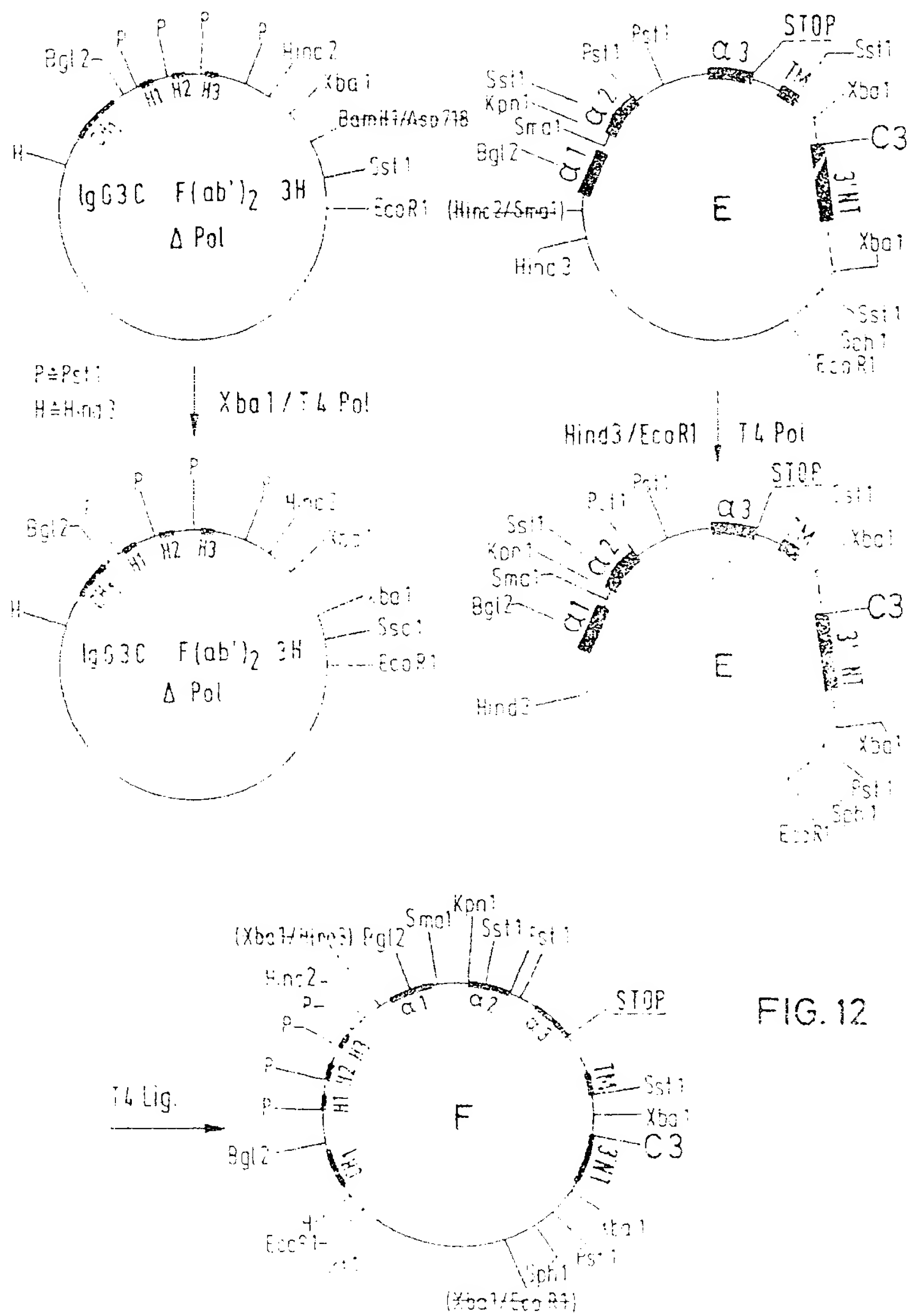
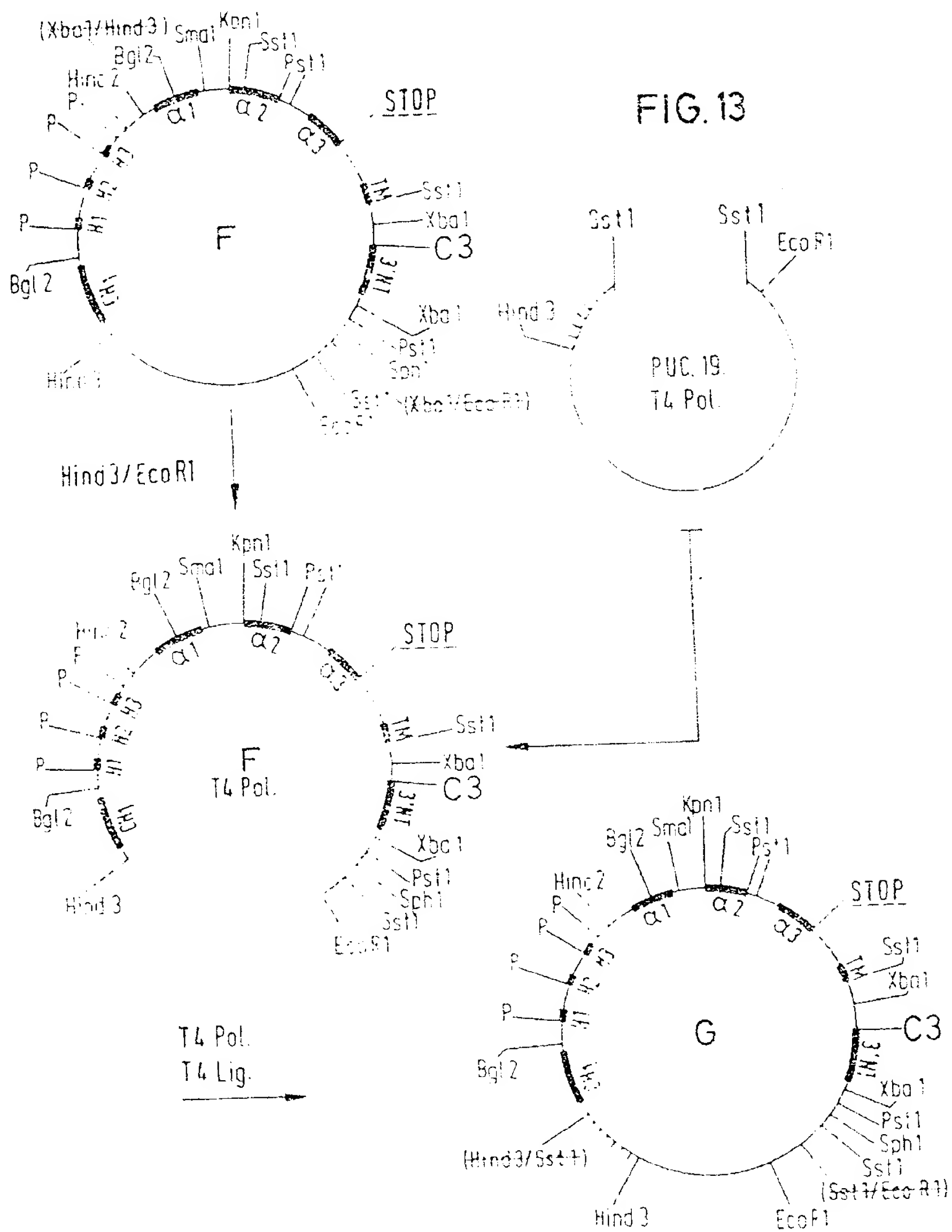


FIG. 12



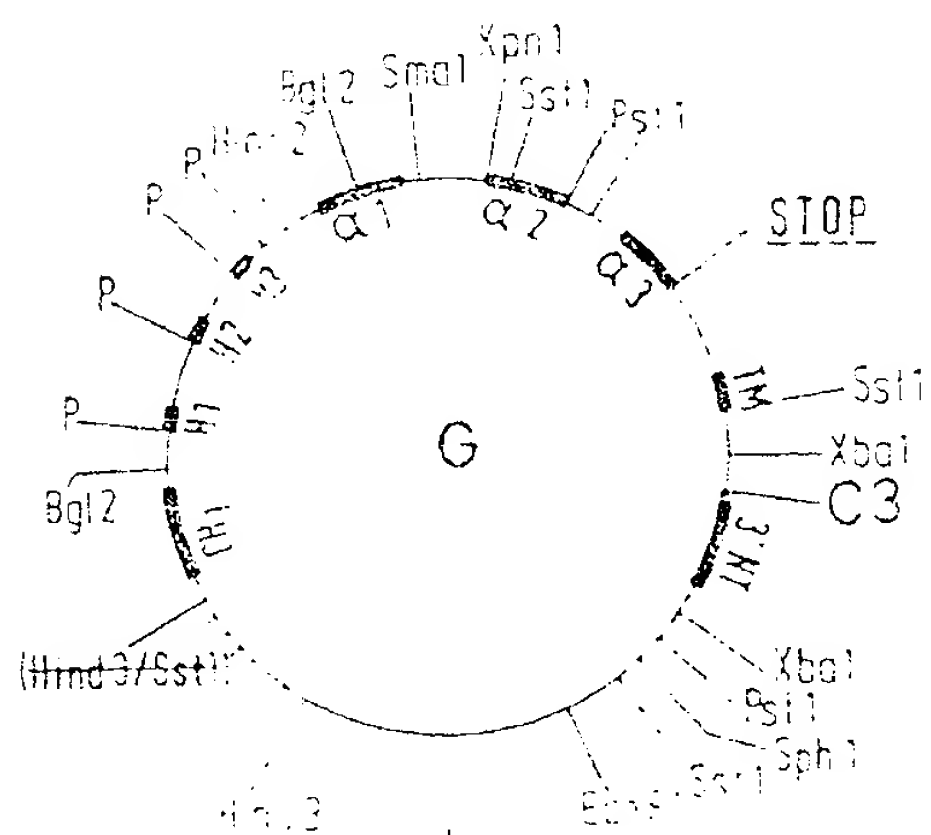
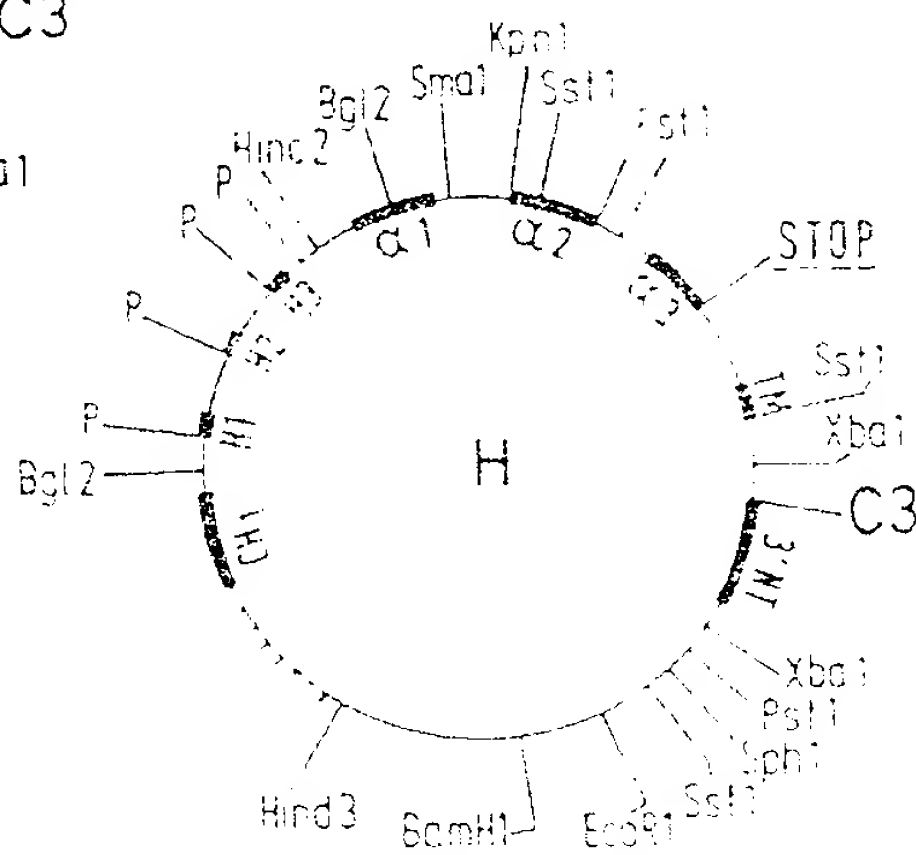
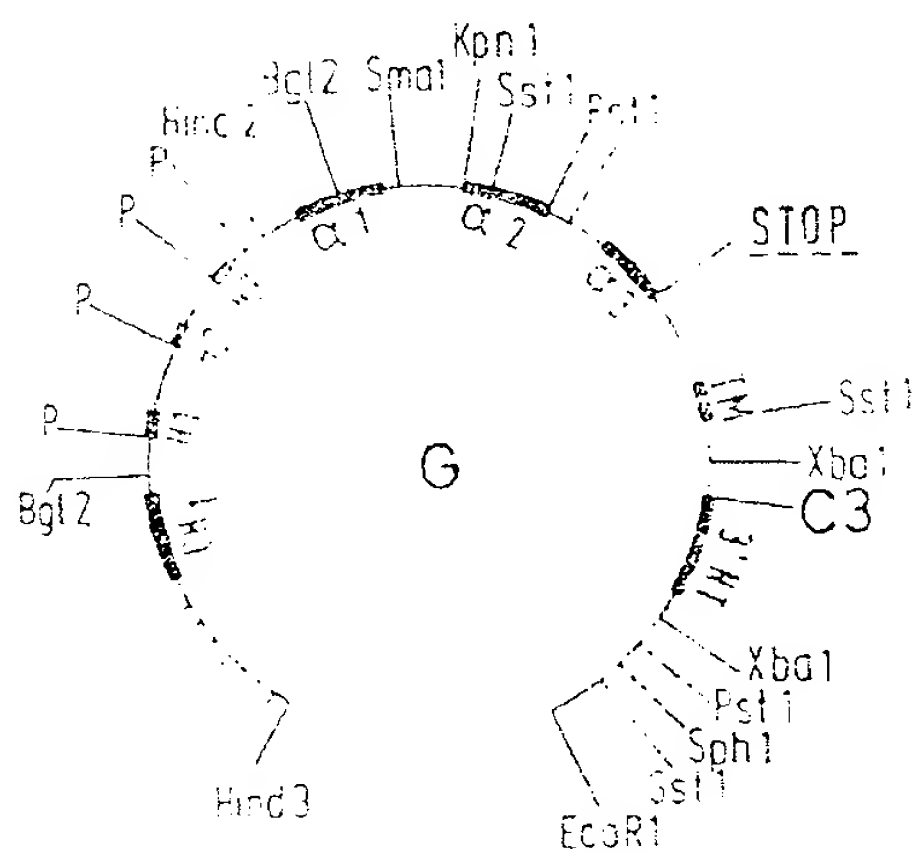
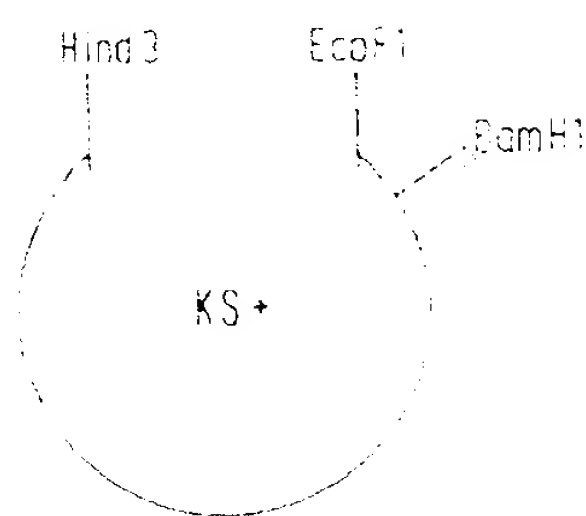


FIG.14



T4 Lig.

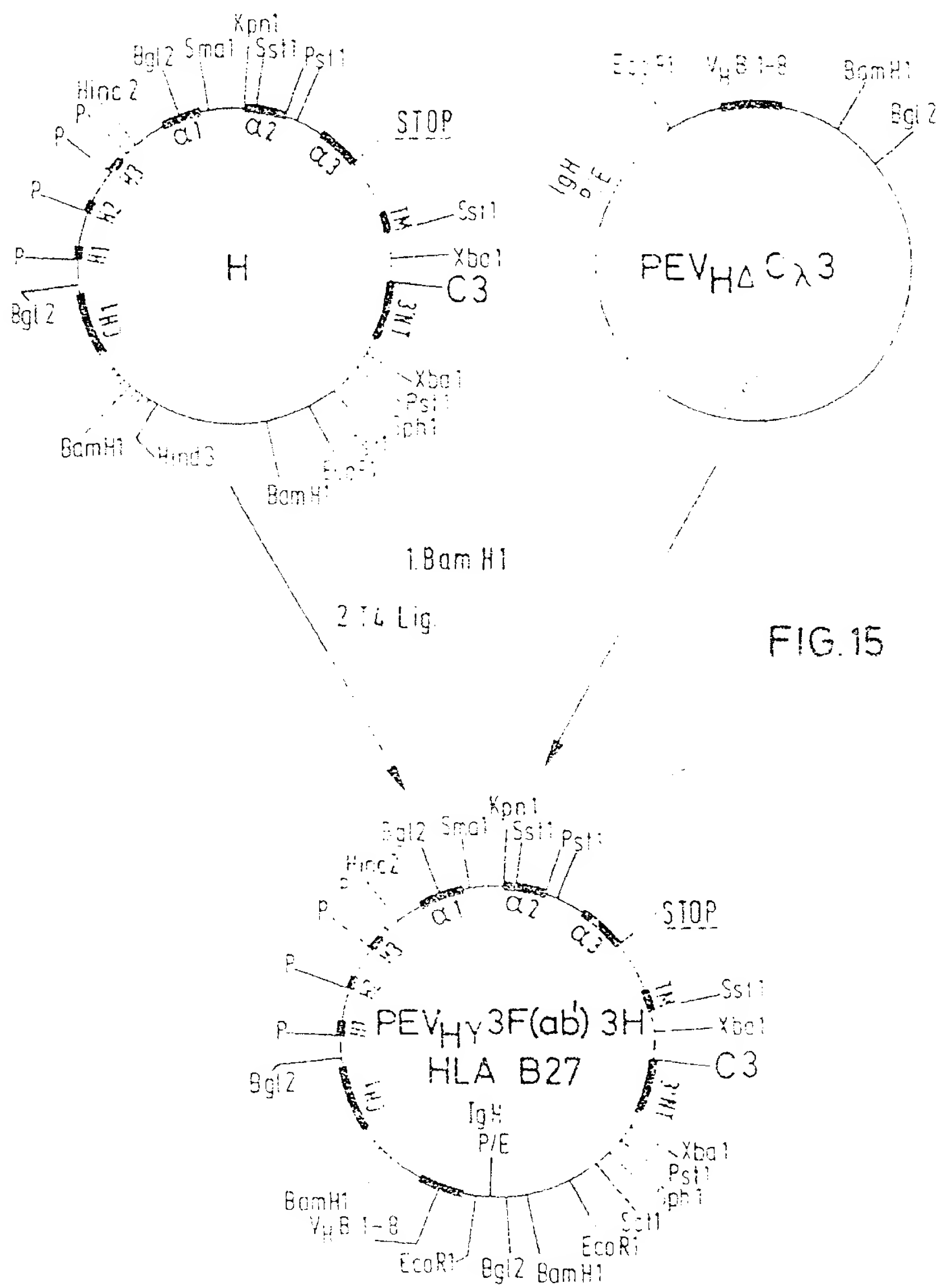
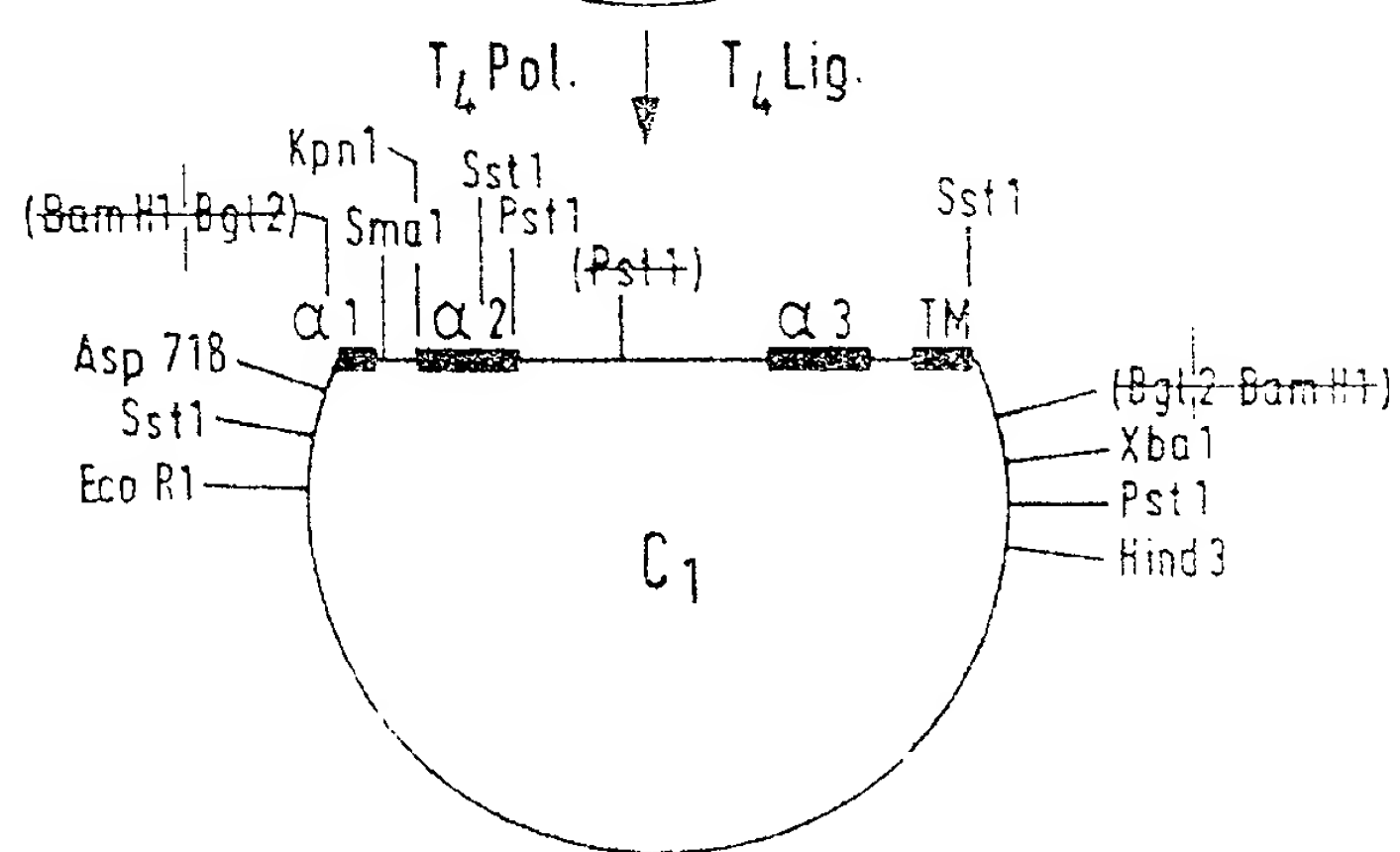
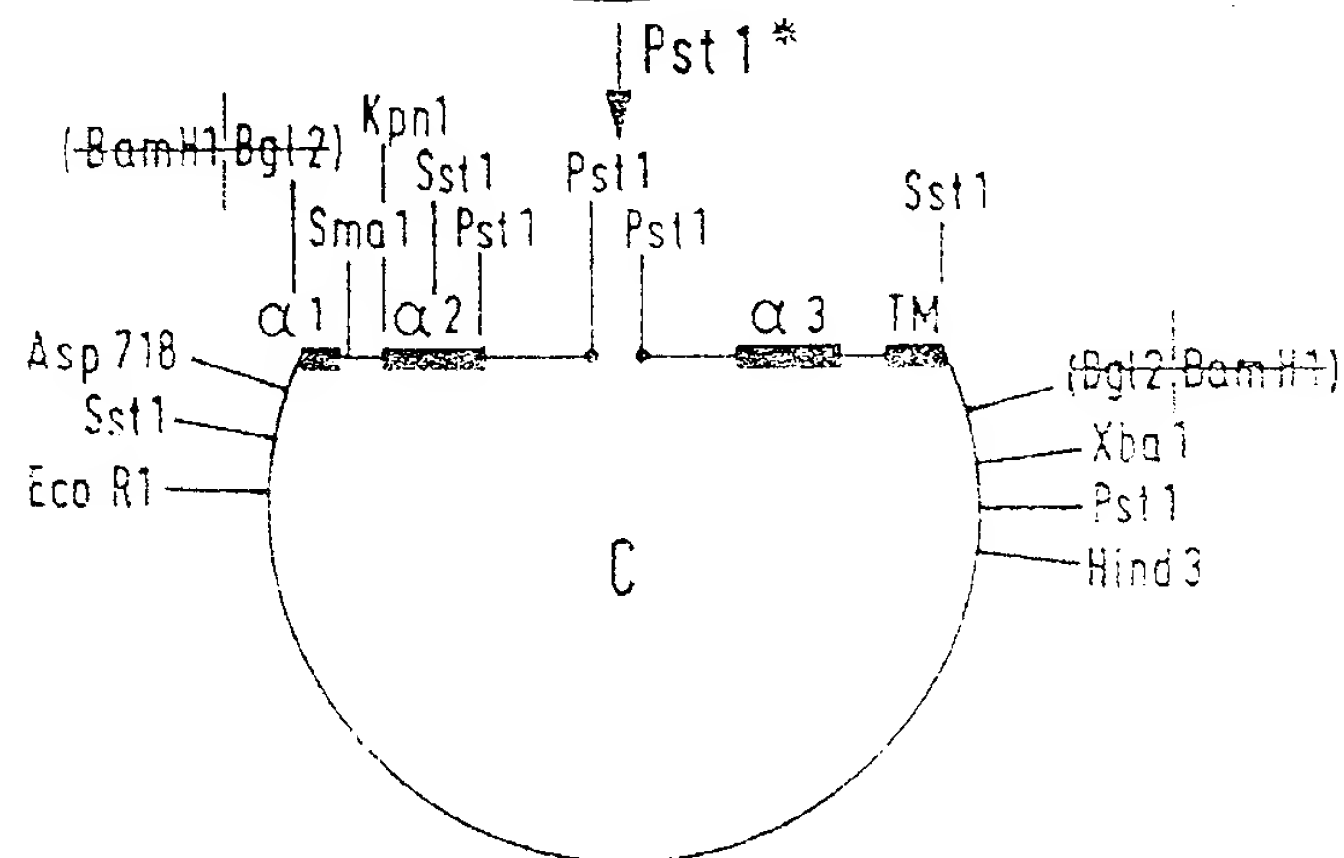
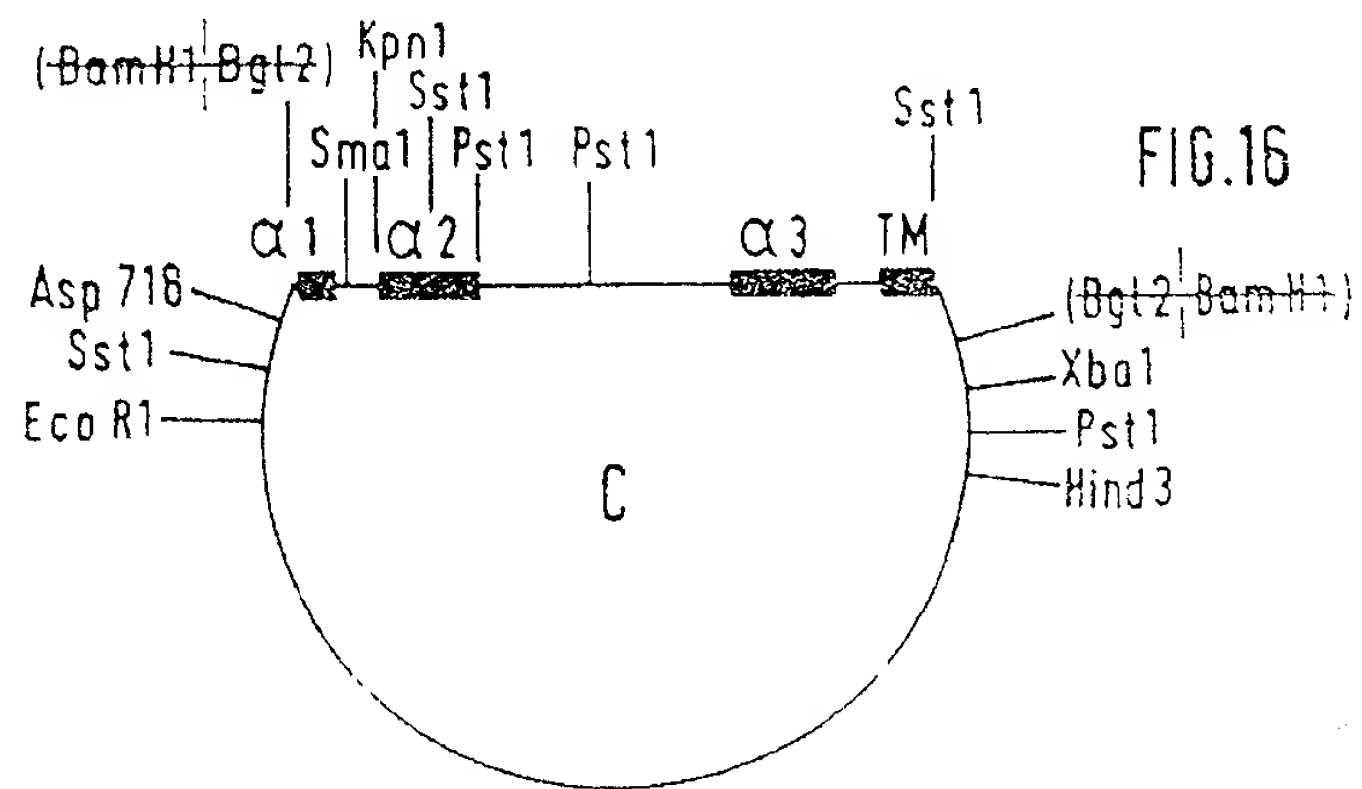


FIG.15



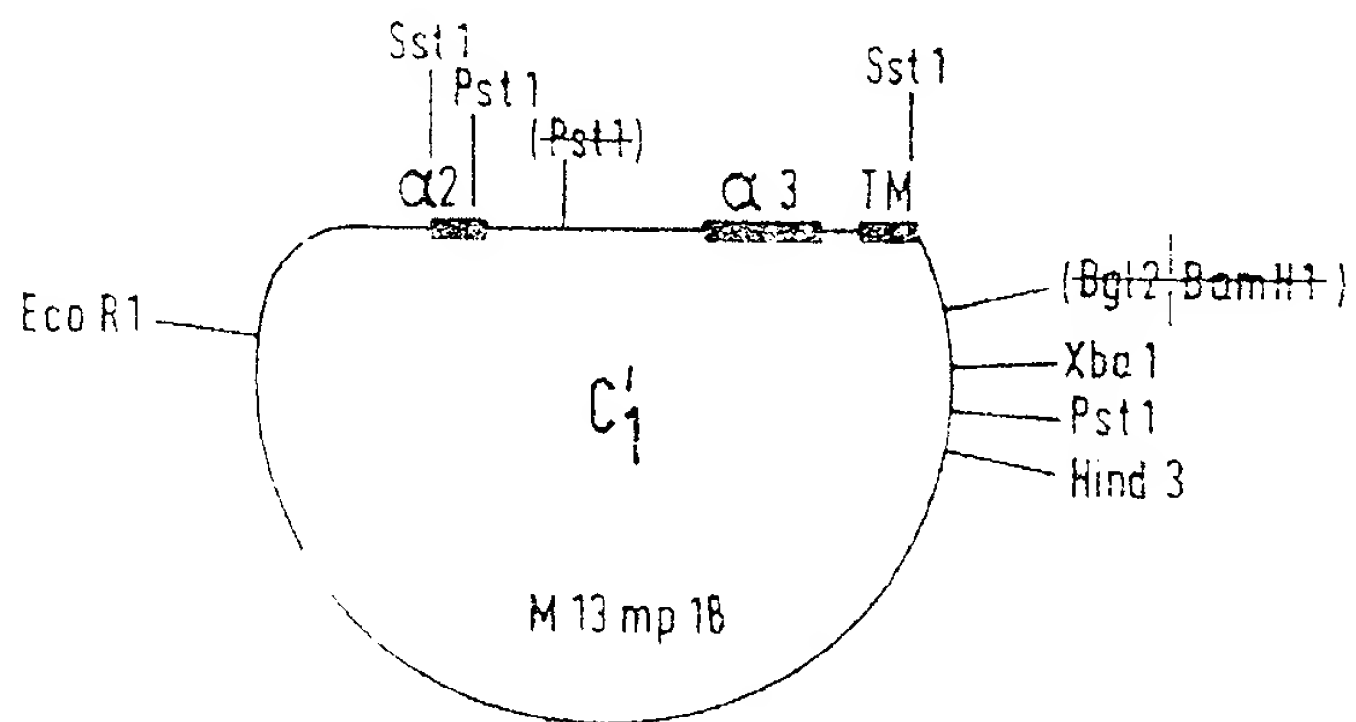
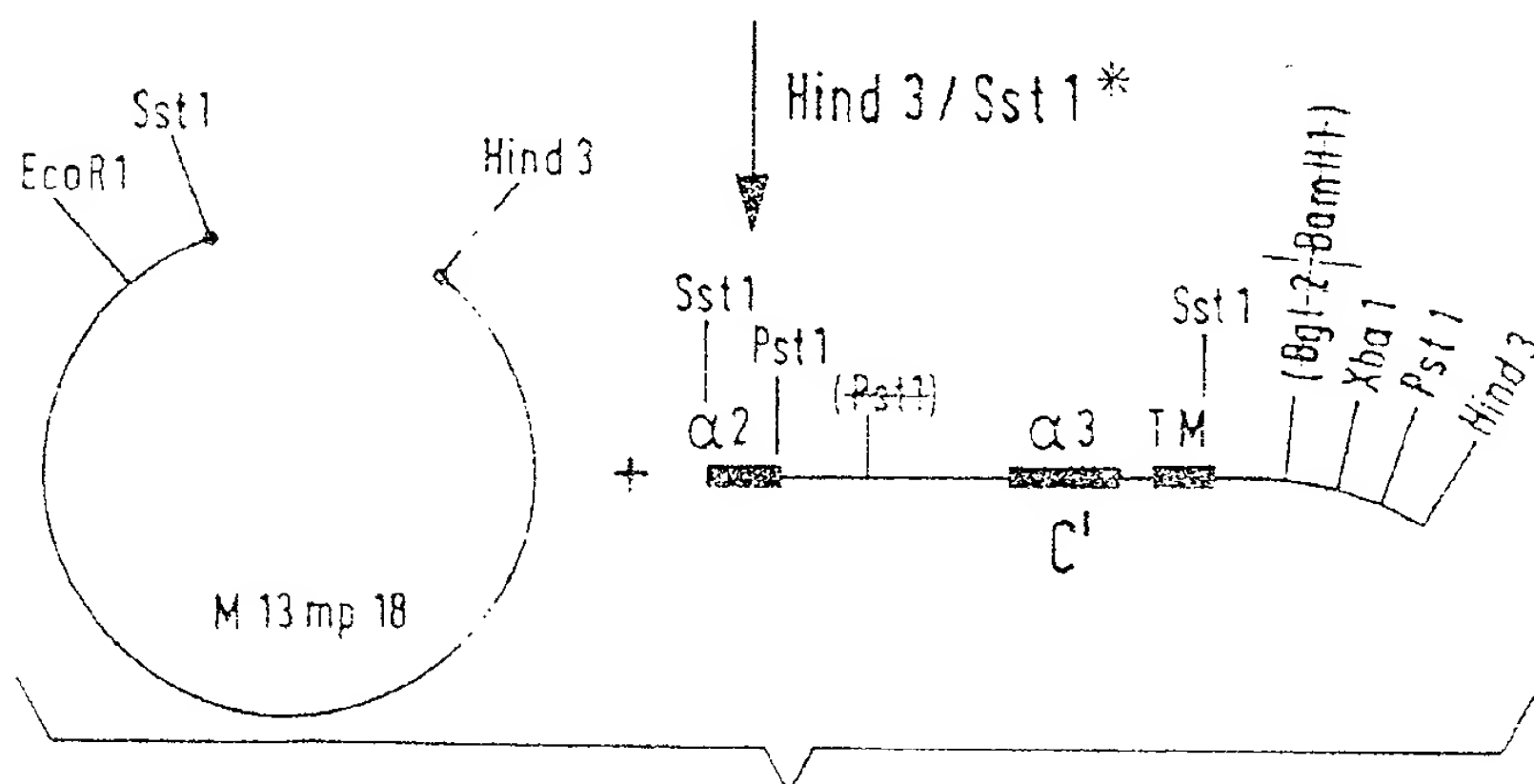
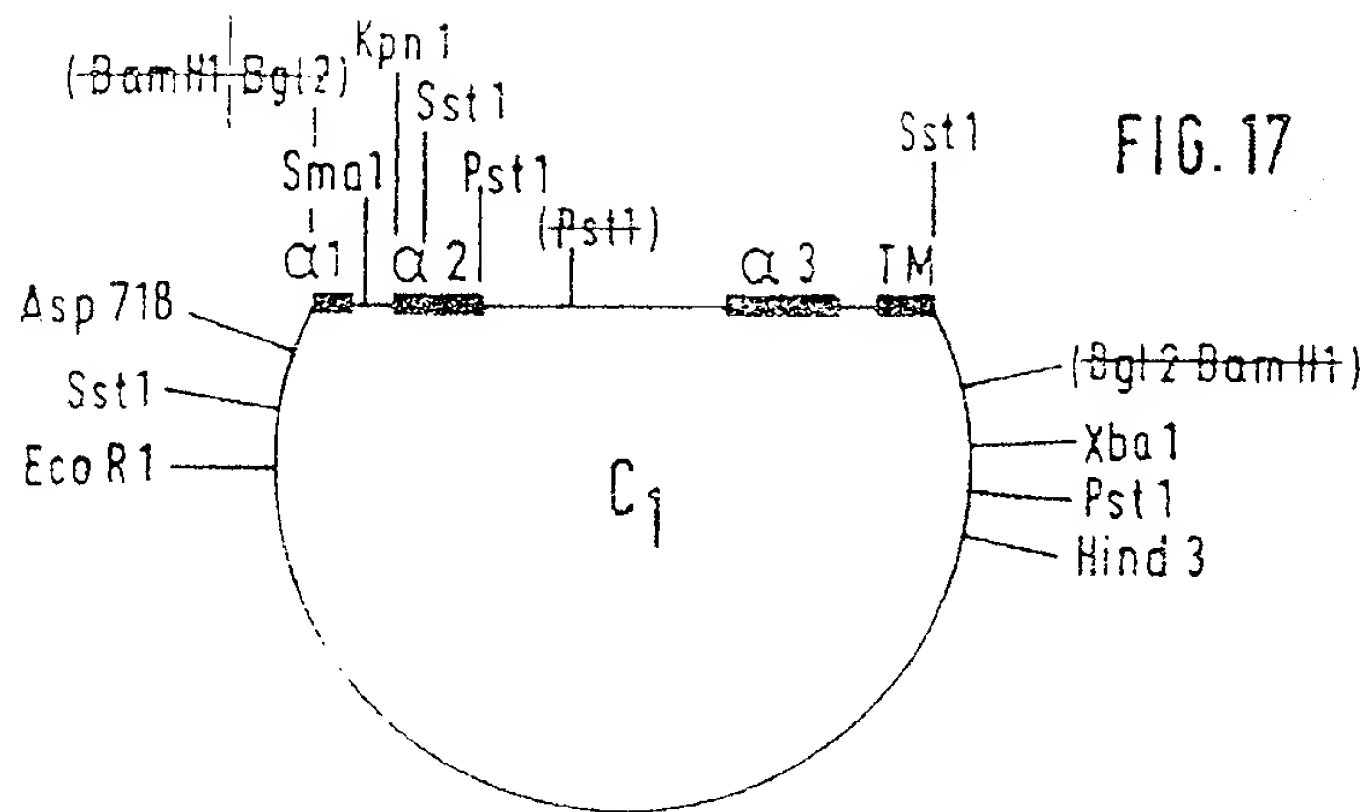


FIG. 18

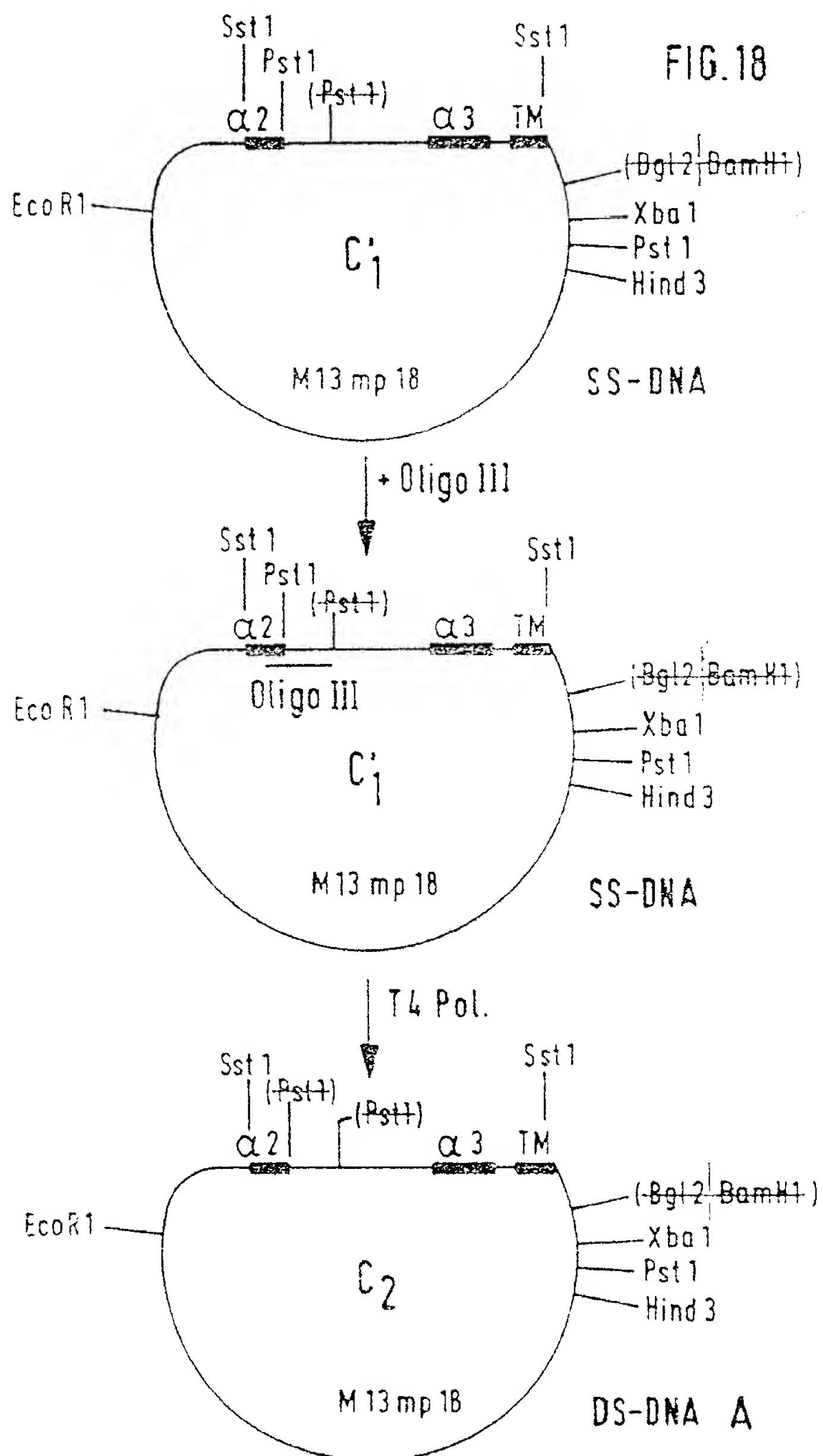
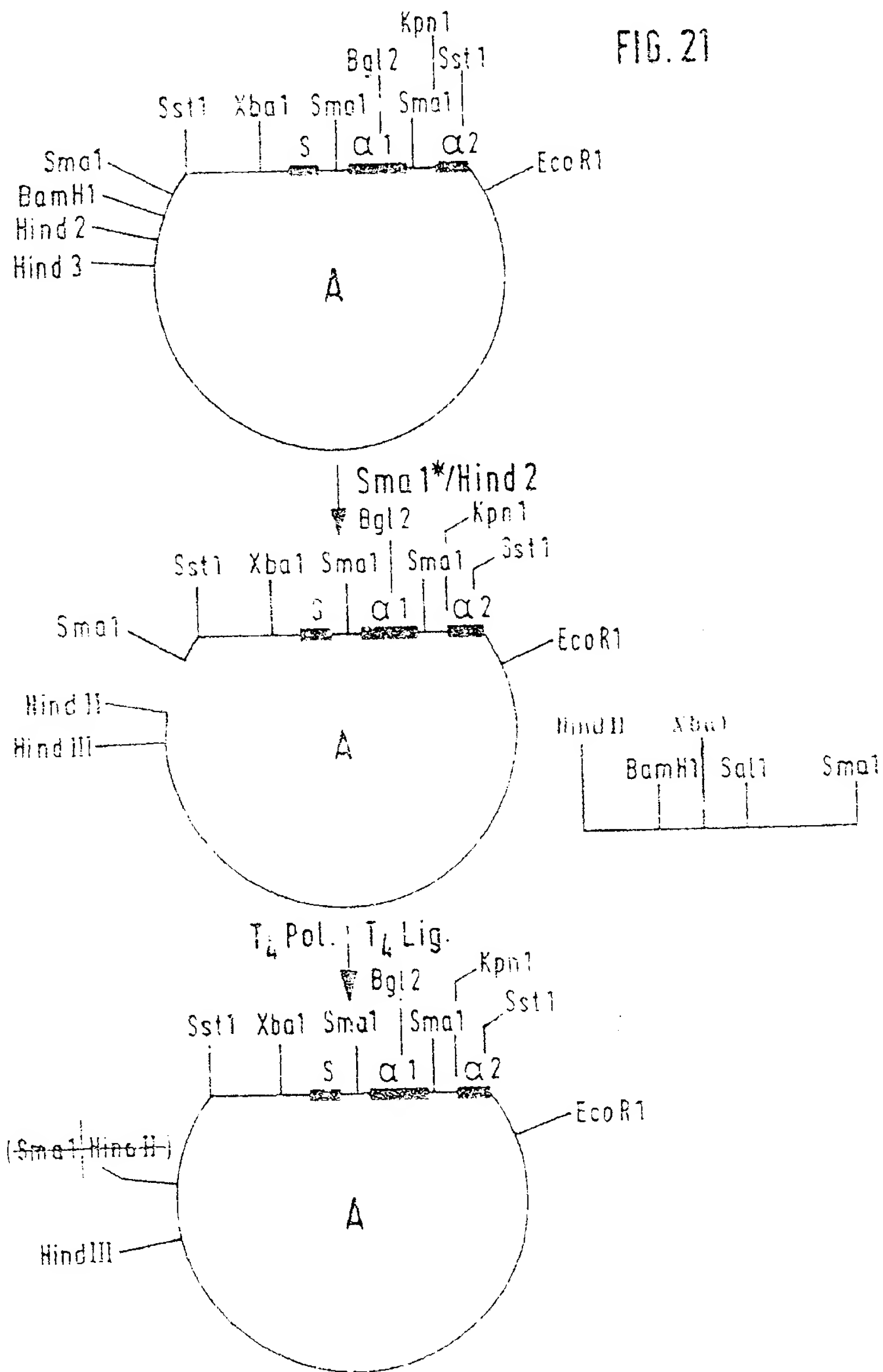
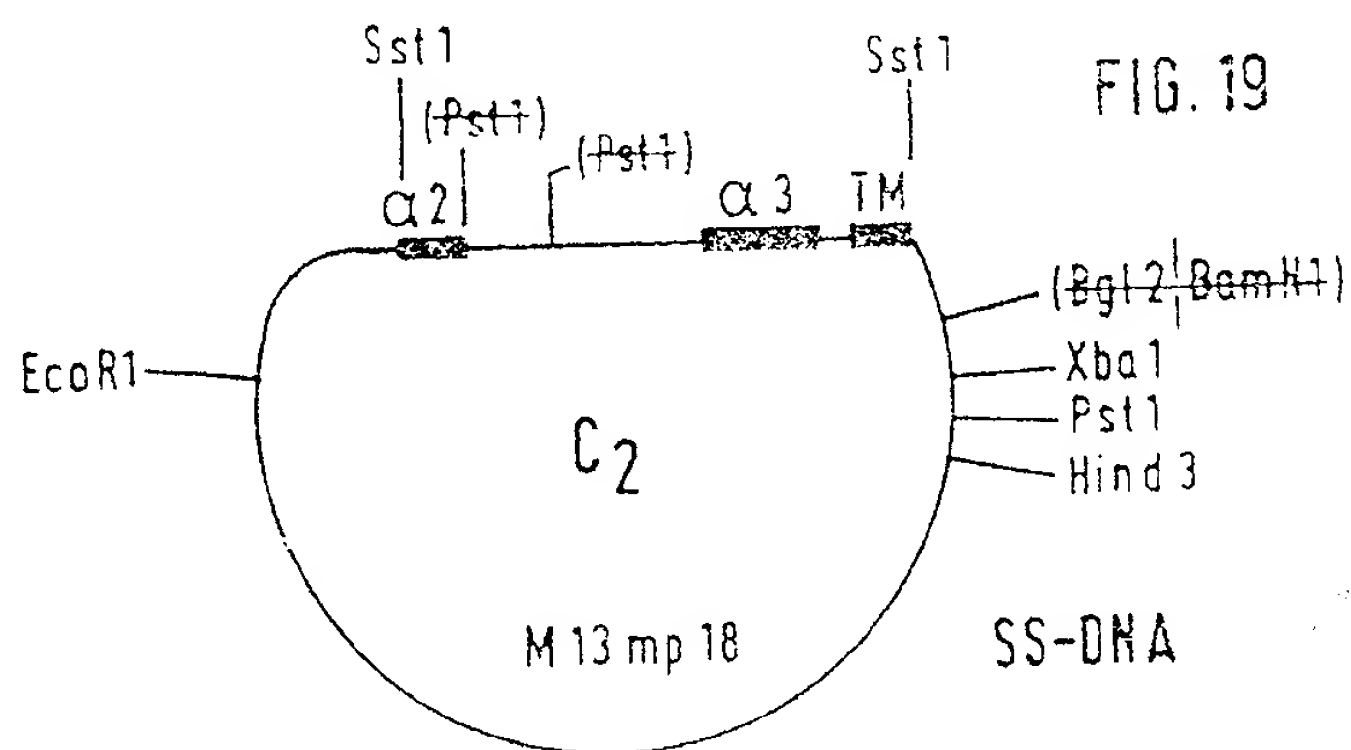


FIG. 21





+ Oligo IV

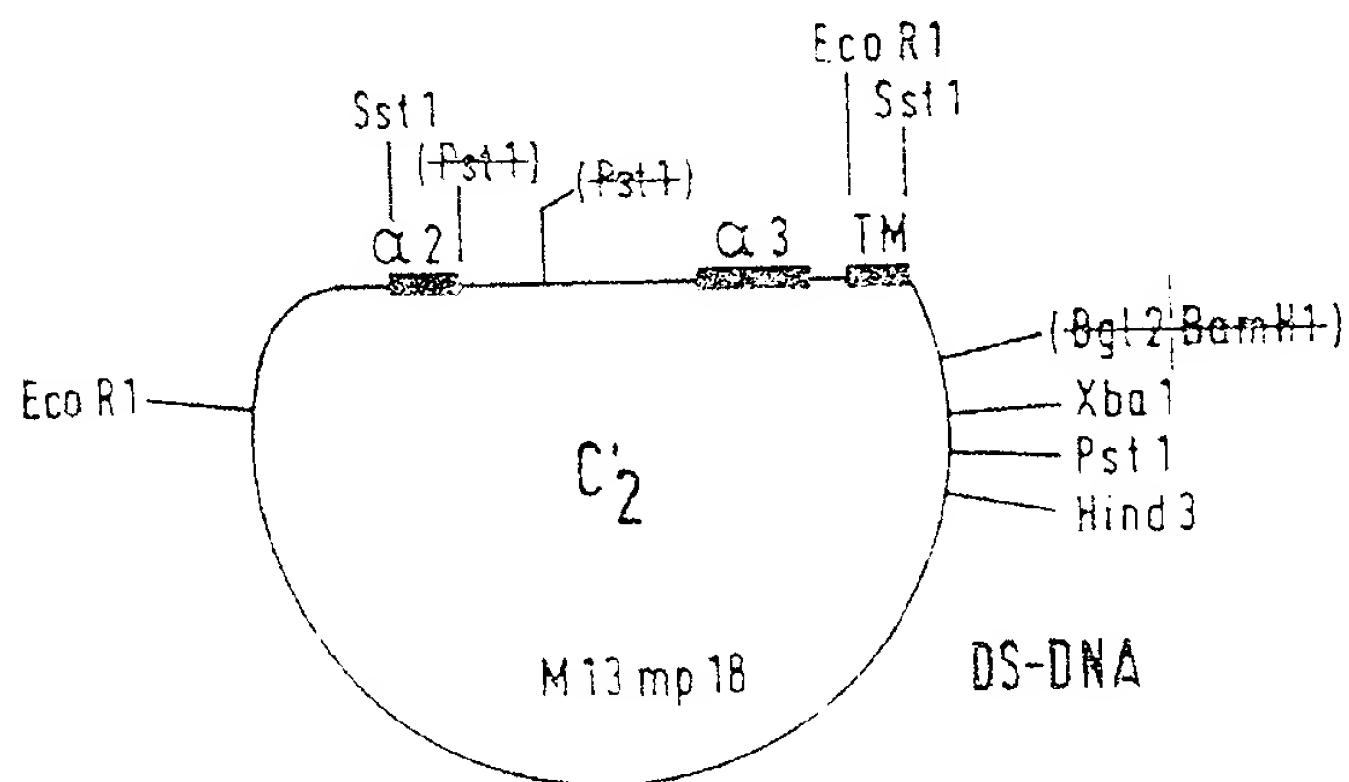
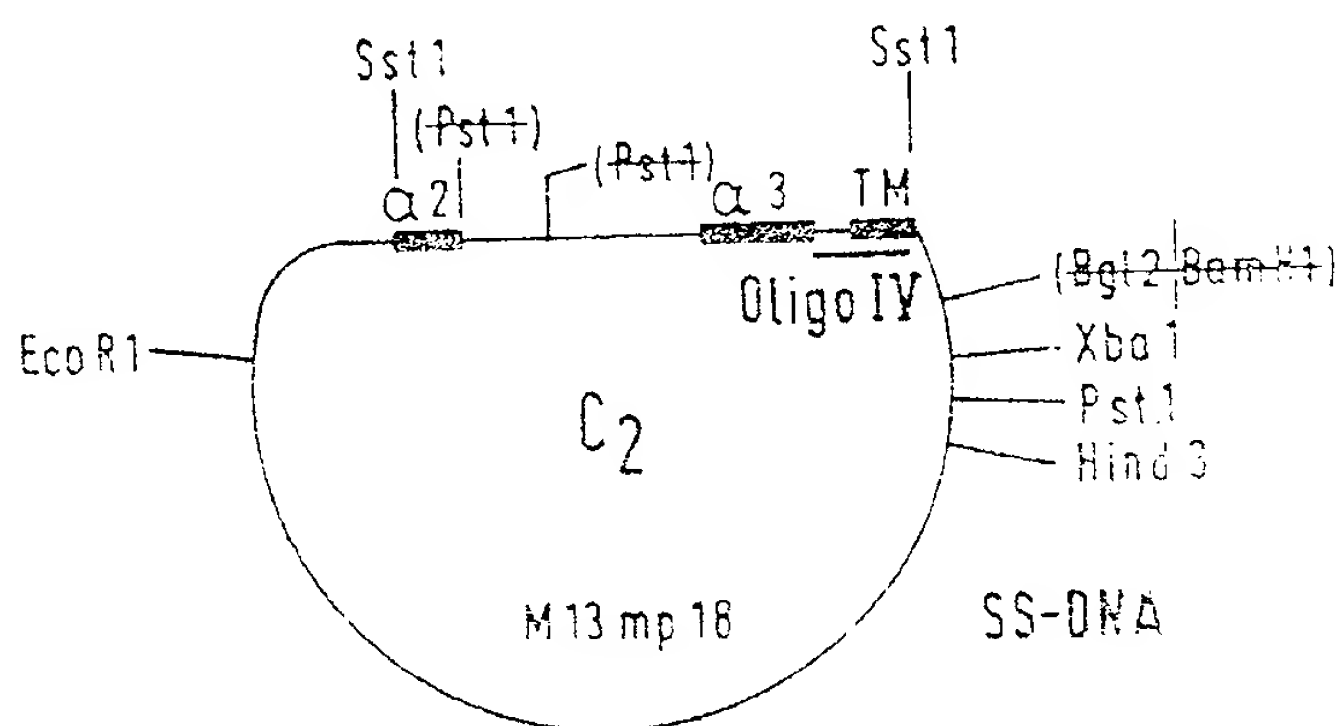


FIG. 28

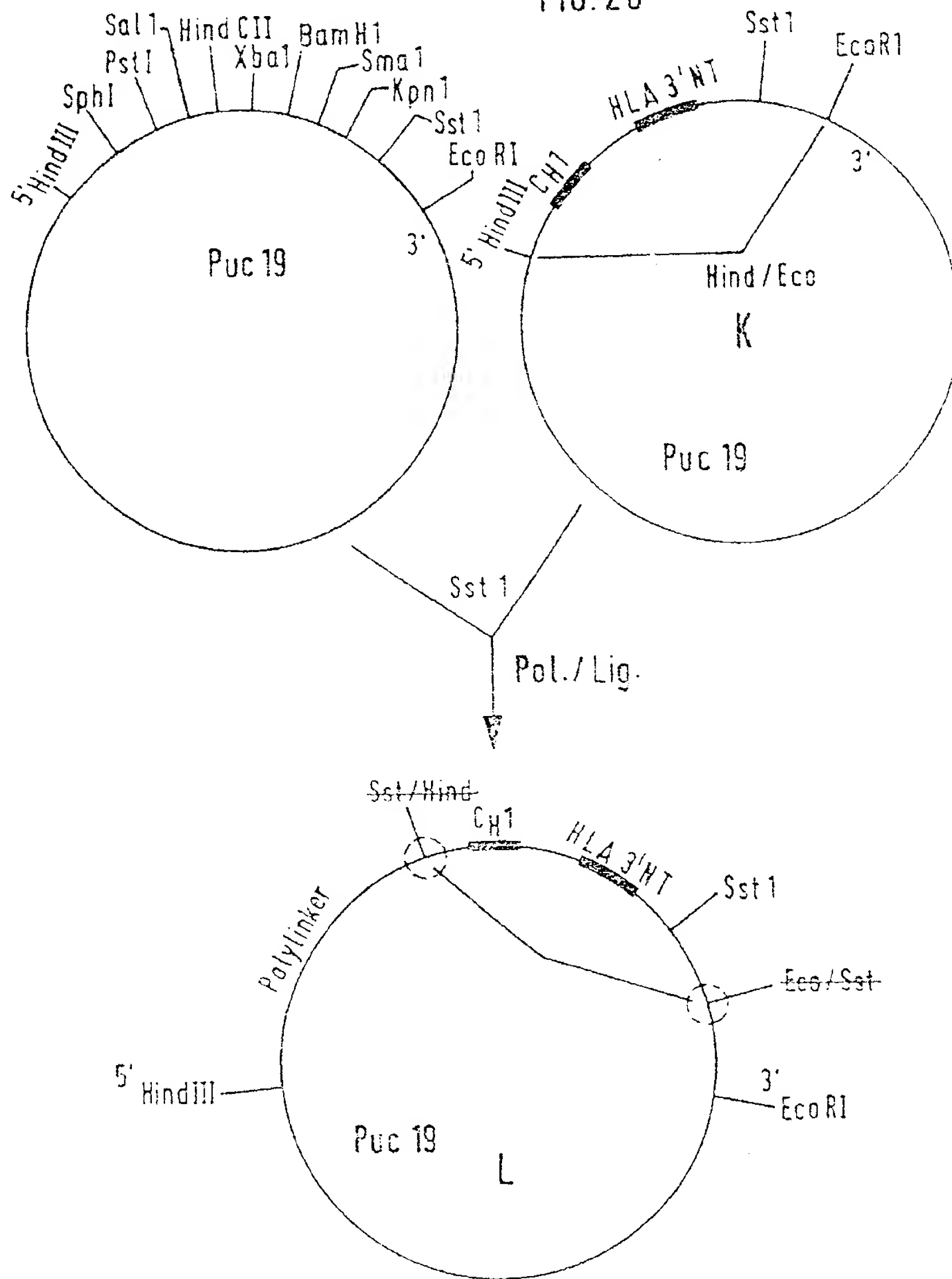
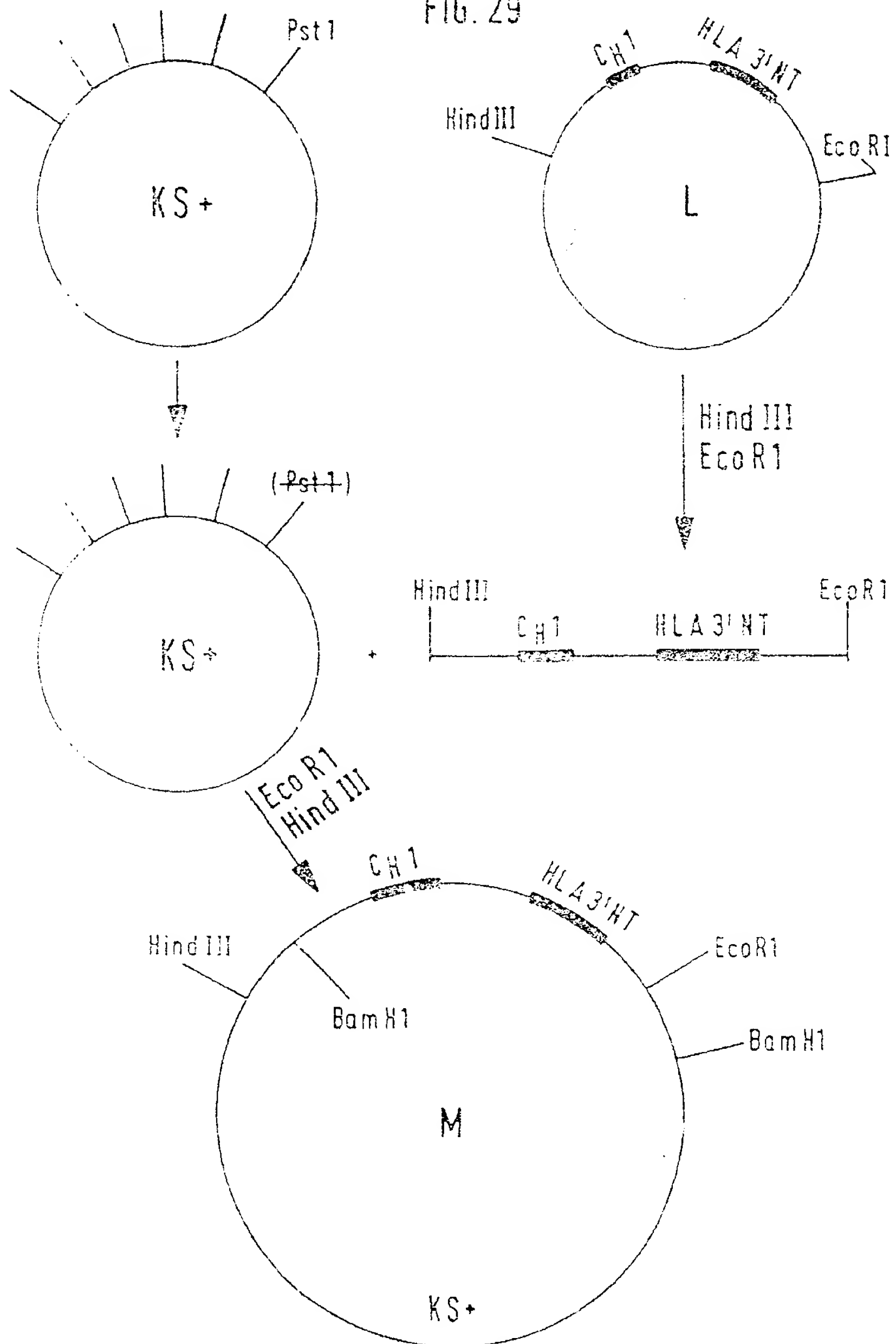


FIG. 29



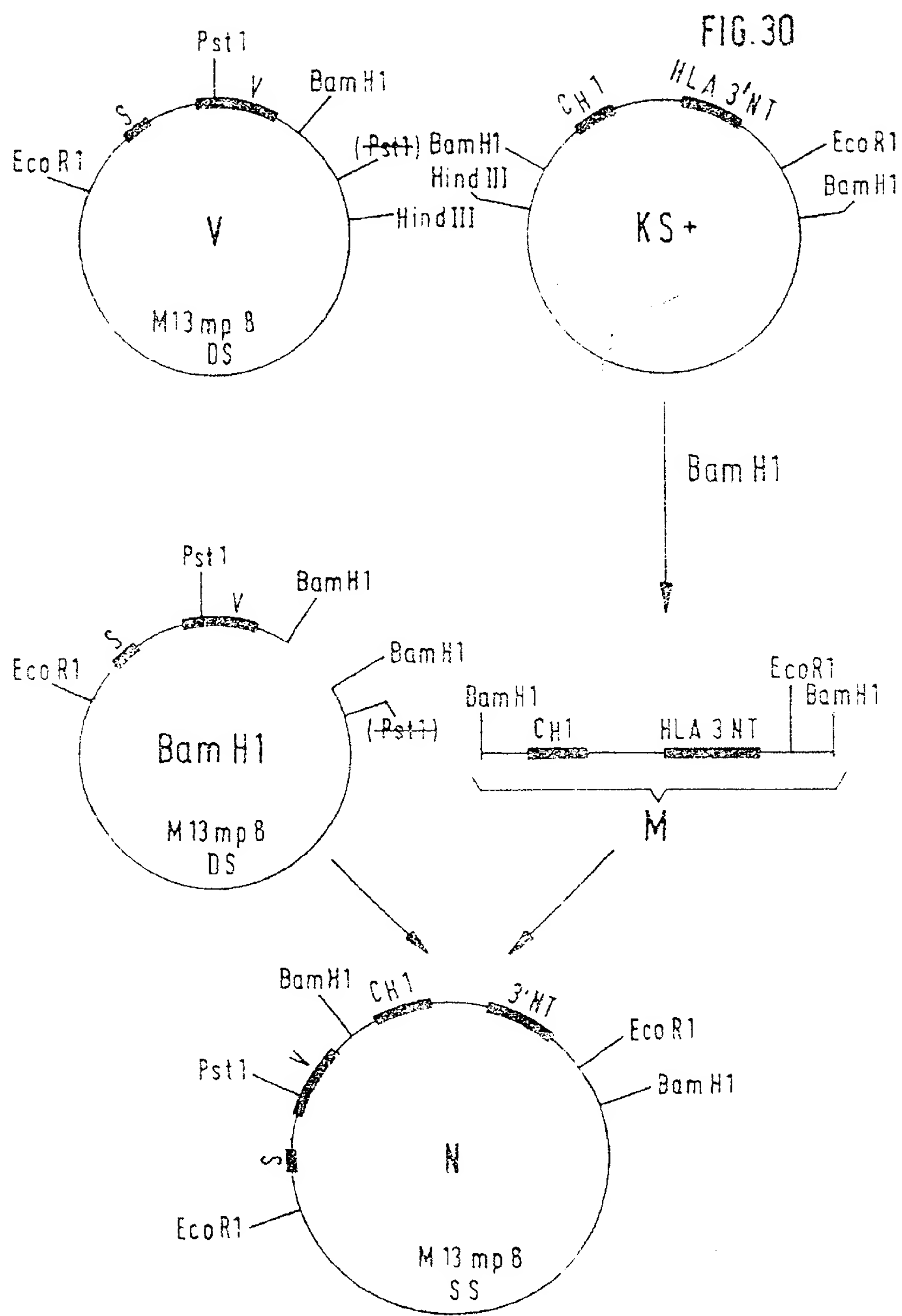


FIG. 31

